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## Heavy metal (PB) bioaccumulation study in *Eisenia fetida* and in the larvae of *Anopheles gambiae* complex using *in silico* drug docking protocols

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### Abstract

Heavy metal bioaccumulation is the process by which the application of *Eisenia fetida* earthworm species in bioremediation of contaminated soil results in a decrease in the concentration of heavy metals. However, it negatively impacts earthworms and hence detrimental to agriculture. On the other hand, in adult mosquitoes, exposure to metal pollution during the larval stage significantly impacted their subsequent insecticide tolerance. The current *in silico* study, we carry out a comparative study on the effect of Heavy metal (PbSO<sub>4</sub>) bioaccumulation in *Eisenia fetida* and in the larvae of *Anopheles gambiae* complex. Metallothionein protein is selected as a common biomarker for the expression of metal accumulation in both species. In order to observe the accumulation of Lead II sulphate in the Metallothionein protein of *Eisenia fetida* and in the mosquito larvae, this study's methodology entails molecular drug docking and H-bond interaction studies using sophisticated automated drug docking servers. The findings unequivocally demonstrate that amino acid residues found in *Eisenia fetida* and in the larvae of the *An. gambiae* complex bind with different heavy metals, most notably Lead II sulphate. Our findings also line up with the previously validated wet lab findings. Finally, it was determined that Lead II sulphate directly binds to Metallothionein amino acid positions of both species. Based on the expression of Metallothionein protein, these results have demonstrated that *Eisenia fetida* and the larvae of the *An. gambiae* complex are capable of accumulating Lead II sulphate in their body. The accumulation of metals in a specific species of earthworm and mosquito larvae, which has detrimental effects on the species' biological systems at high concentrations, is clearly described throughout this entire *In silico* study.

**Keywords:** Lead II sulfate, *Eisenia fetida*, larvae of *An. gambiae* complex, docking

### 1. Introduction

Heavy metal pollution is currently a major problem in many parts of the world <sup>[1, 2]</sup>. Chemically, heavy metal elements, such as mercury (Hg), cadmium (Cd), copper (Cu), nickel (Ni), lead (Pb), arsenic (As), chromium (Cr), and zinc (Zn), have an atomic mass greater than 20 and a gravity greater than 5 g•cm<sup>-3</sup> <sup>[3, 4, 5]</sup>. Because of its similar chemical properties and environmental response, metalloid arsenic is often classified as a heavy metal <sup>[7, 8]</sup>. Heavy metal contamination of agricultural soil has been a major concern in China ever since industrialization and technological advancement began to take hold <sup>[9]</sup>. China has serious soil metal pollution, according to the State Environmental Protection Administration <sup>[10]</sup>. Among all the contaminants, environmental chemists have focused particularly on heavy metals because of their toxicity and persistence.

Toxic hazardous materials have been polluting soil all over the world for the past few decades, which is a major cause for concern <sup>[11, 12]</sup>. Hard to decompose, soil hazard elements can find their way into water and food supply networks, posing a long-term threat to human health and food safety <sup>[13, 14]</sup>. The United States Environmental Protection Agency (USEPA) has designated heavy metals as priority control contaminants, and because of their potentially dangerous, chronic, and irreversible characteristics, they are receiving more and more attention globally <sup>[15, 16]</sup>. Although too much copper is harmful to the body, it is a necessary trace element <sup>[17]</sup>.

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Pb is the second most common heavy metal in the environment, after Cd, according to two recent surveys conducted by Chinese government ministries [18]. According to data from China's prestigious National Soil Pollution Investigation, 1.50 percent of soil samples had Pb contamination from 2005 to 2013 [19,20]. Humans are primarily exposed to lead through hand-to-mouth contact with dust and soil, especially in young children. [21] Blood lead poisoning is a serious public health issue, particularly in developing countries [22]. Children that reside close to the smelters have elevated blood lead levels (BLLs) [23].

Our attention is directed towards the build-up of Lead II sulphate (PbSO<sub>4</sub>) in *Eisenia fetida* via soil and in the larvae of *Anopheles gambiae* complex. We determine how Lead II sulphate inhibits the expression of the protein, Metallothionein through Molecular Drug Docking studies. Waste from industry, agriculture, and cities has been successfully disposed of using the main earthworm species: *Eisenia foetida*, *E. andrei*, *E. eugeniae*, and *Perionyx excavates*. [24] *Erythra fetida* is widely used in earthworm composting due to its ease of cultivation. Information regarding its biological traits is abundant. *E. fetida* is capable of producing earthworm biomass and is also capable of decomposing organic waste and releasing wormcast [25].

*Anopheles gambiae* species complex members generally favour breeding in open, sunny, temporary, mostly unpolluted bodies of water [26]. As a result, information about members of this complex reproducing in contaminated water indicates a substantial biological change in this species [27, 28, 29]. Moreover, this transition has been documented in members of this species complex, such as *An. arabiensis*, in addition to *An. gambiae sensu stricto* [30, 31]. This adaptation may unintentionally alter biological traits of epidemiological significance, such as insecticide susceptibility, since non-pesticidal residues have been shown to modify detoxification enzyme capacity [32, 33, 34] and insecticide resistance phenotypes [35].

## 2. Methodology

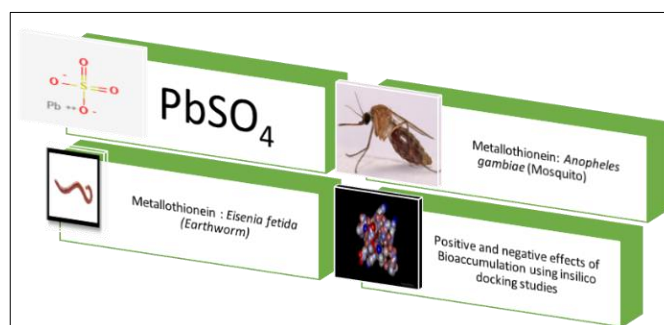
The methodology in this research study includes: 1. Target Sequence Selection. 2. Drug Docking Studies. 3. Visualization of 3D Protein – Metal Binding Interaction. (Fig :1)

**1. Target Sequence Selection:** The NCBI (National Centre for Biotechnology Information) [36] <https://www.ncbi.nlm.nih.gov/> database was used to identify the potential (PbSO<sub>4</sub>) metal interacting protein sequence, Metallothionein (MT\_EISFE) in *Eisenia fetida* (AUS83918.1) and in (AAX86007.1) *Anopheles gambiae* because the database has the sequence available, based on the sequence that has been experimentally proven and the related references. [37] This sequence was mainly retrieved in order to dock it with Lead (II) Sulfate (PbSO<sub>4</sub>) metal and to validate the efficiency of the interactions.

**2. Drug Docking Studies:** Numerous studies in the literature have demonstrated that lead (II) sulphate is present in soil [38]. Hence Lead (II) Sulfate (CID: 24008) was selected using NCBI PubChem Compound Database (<https://pubchem.ncbi.nlm.nih.gov/>). Metallothionein of *Eisenia fetida* and *Anopheles gambiae* was introduced to Lead (II) Sulfate and the metal binding to the structural domain regions of the protein sequence was viewed using HDock server [39] <http://hdock.phys.hust.edu.cn/>.

**3. Visualization of 3D Protein - Metal Binding Interaction:** Discovery Studio, a molecular visualisation

tool, was used to validate the docking results. This software facilitates the visualisation of the intramolecular interactions between *Eisenia fetida*'s MT protein and lead (II) sulphate and between *An. gambiae*'s MT protein and lead (II) sulphate.

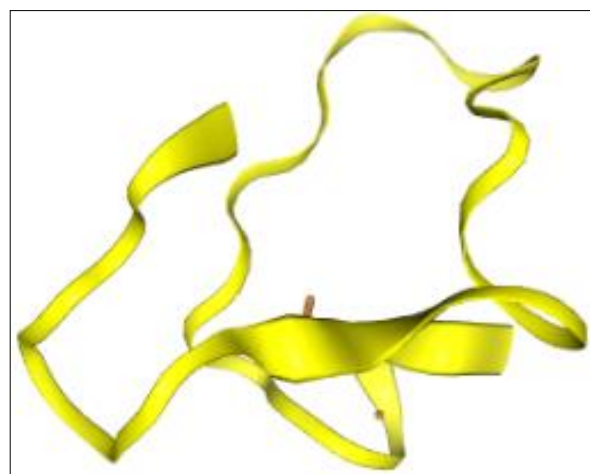


**Pictorial Representation of the Research Methodology**

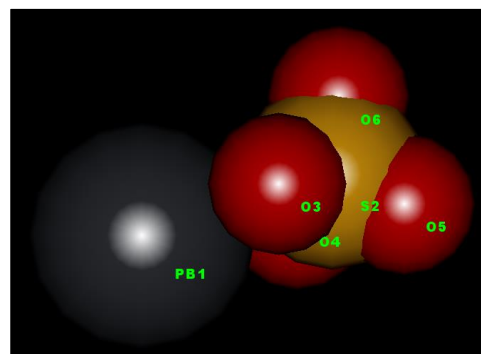
## 3. Results

```
>AUS83918.1 Metallothionein [Eisenia fetida]
MADALDTQCCGKSTCAREGSTCCCTNCRCLKSECLPG
CKKLCADAEEKGKCGNAGCKCGAACKCSAGSCA
AGCKKGCCGD
```

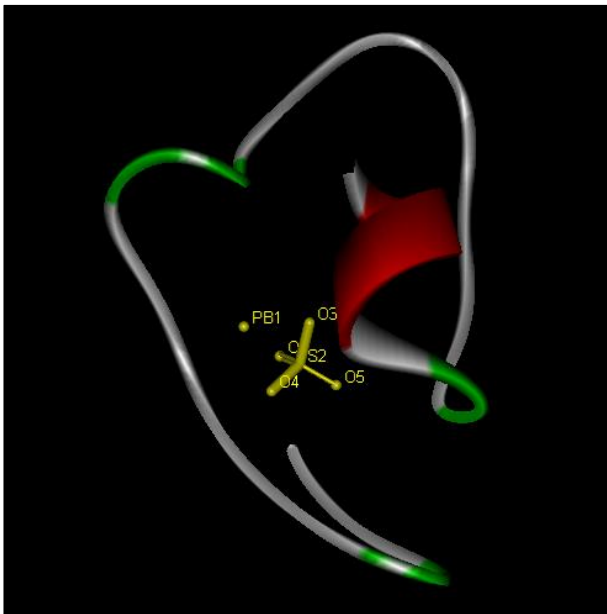
**Fig. 1:** FASTA format of the Metallothionein sequence of *Eisenia fetida* and its corresponding amino acid sequence



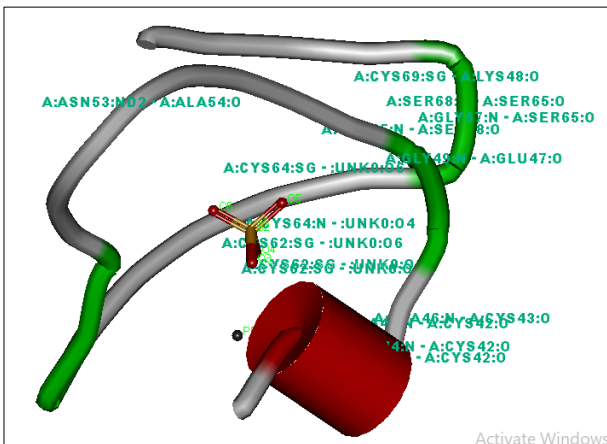
**Fig 2:** 3D structure of Metallothionein of *Eisenia fetida* viewed using Discovery Studio software



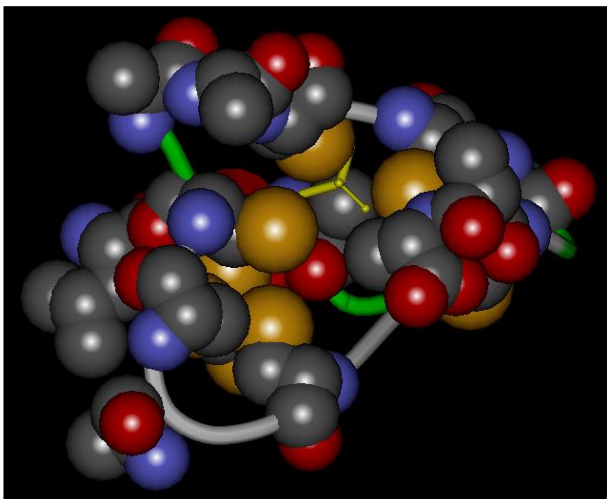
**Fig 3:** 3D structure of PbSO<sub>4</sub> viewed using Discovery Studio software in space fill model with respective coloured atomic labels.



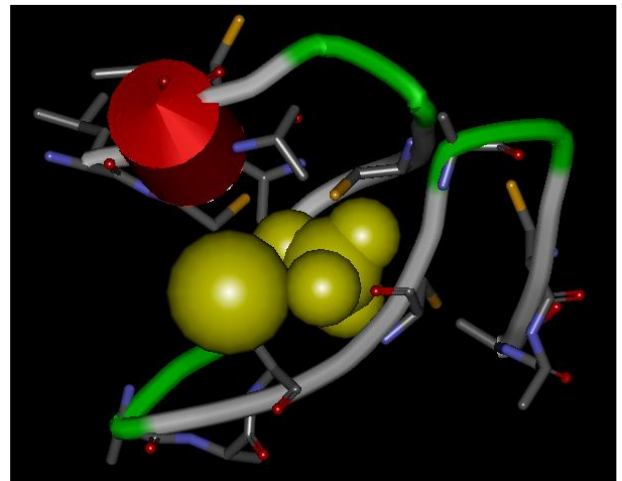
**Fig 4:** Complex form of Lead II sulphate and Metallothionein of *Eisenia fetida* viewed using Discovery studio software



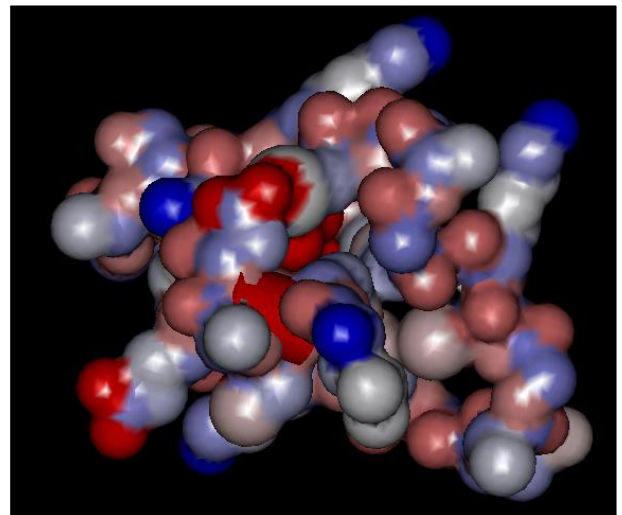
**Fig 6:** 3D structure complex form of Lead II sulfate and Metallothionein (*Eisenia fetida*) with respective hydrogen bond amino acid labels viewed using Discovery Studio software.



**Fig 8:** 3D structure complex form of Lead II sulfate and Metallothionein (*Eisenia fetida*) with respective hydrophobic interaction sites viewed using Discovery Studio software.



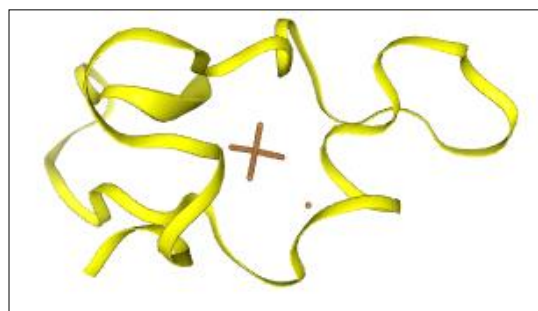
**Fig 5:** 3D structure complex form of Lead II sulfate (represented in yellow colour spacefill model) and Metallothionein (*Eisenia fetida*) with respective hydrophobic interaction sites viewed using Discovery Studio software.



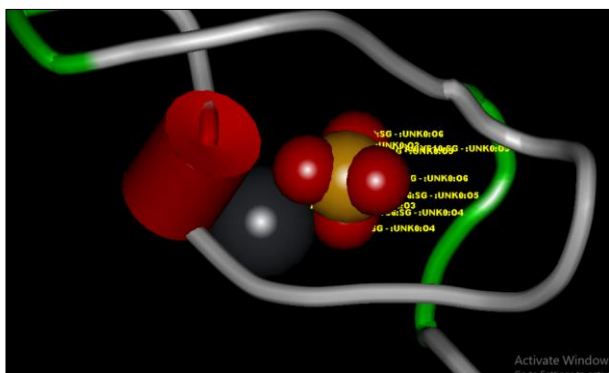
**Fig 7:** 3D structure complex form of Lead II sulfate and Metallothionein (*Eisenia fetida*) with electrostatic interactions between them viewed using Discovery Studio software.

>AAX86007.1 Metallothionein 2, partial [*Anopheles gambiae*]  
MPCKTCVADCKCTSPNCGAGCGCESRCTPCCKDGAKE  
GCC

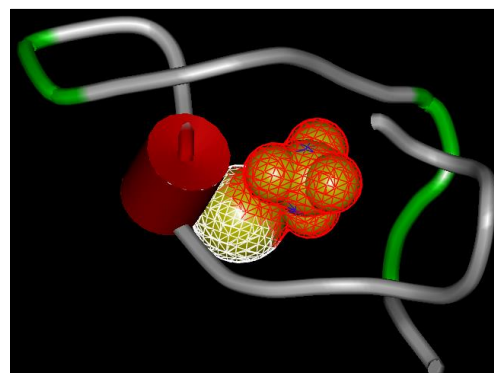
Fig: 9 FASTA format of the Metallothionein sequence of *An.Gambiae* and its corresponding amino acid sequence



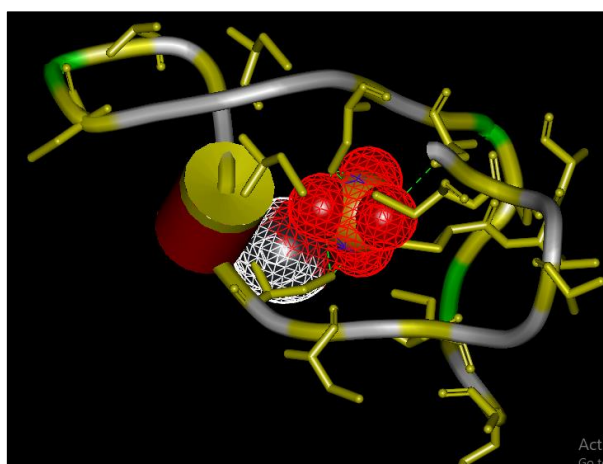
**Fig 10:** 3D structure of Metallothionein of *An.Gambiae* viewed using Discovery Studio software.



**Fig 11:** Complex form of Lead II sulphate and Metallothionein of An.Gambiae viewed using Discovery studio software with respective binding amino acids.



**Fig 12:** Complex form of Lead II sulphate and Metallothionein of An.Gambiae viewed using Discovery studio software showing 3D electrostatic interaction force



**Fig 13:** Complex form of Lead II sulphate and Metallothionein of An. Gambiae viewed using Discovery studio software showing 3D electrostatic - hydrophobic interaction force

**Table 1:** Drug docking summary

|   | <b>Protein Target 1</b>                                | <b>Protein Target 2</b>                                   |
|---|--|---|
| <b>Heavy Metal Compound 1</b>                     | Metallothionein [ <i>Eisenia fetida</i> ] (AUS83918.1) | Metallothionein [ <i>Anopheles gambiae</i> ] (AAX86007.1) |
| Lead II Sulfate (PbSO <sub>4</sub> ) (CID: 24008) | -35.39 kcal/mol.                                       | -41.39 kcal/mol.  |

Table 1 Molecular binding affinity scores with units between the Metallothionein *Eisenia fetida* and *Anopheles gambiae* heavy metal (PbSO<sub>4</sub>)

**4. Discussion**

In this study, the length of the Metallothionein protein sequence of *Eisenia fetida* is 80 aa amino acids (Fig: 2, 3), and the length of the Metallothionein protein sequence of *An. Gambiae* is (41 aa) (Fig: 9, 10) that of the selected heavy metal, Lead (II) sulphate (PbSO<sub>4</sub>) (Anglislite) (PubChem CID: 24008) is 303.92838 g/mol molecular weight. It contains Topological Polar Surface Area of 88.6 A<sup>2</sup> and a heavy atom count of 6. (Fig: 3) An important part of improving the biological, chemical, and physical characteristics of soil is done by earthworms. They serve as major bioindicators of environmental contamination and are regarded as keystone species within ecosystems.

Agriculture has been based on soil for ages, as soil is the most valuable natural resource and mankind's greatest legacy. As a result of human progress towards industrialization, dangerous pollutants like heavy metals, toxins, and carcinogenic

compounds are produced and released into the environment. It is well known that earthworms have positive effects on plant growth, nutrient cycling, and soil fertility. By burrowing and casting, earthworm activity improves the physical conditions of the soil by forming stabilised aggregates that facilitate easy air and water penetration. Because of their toxicity and tendency to accumulate in the environment, heavy metals are the primary environmental pollutants and a major concern. Heavy metal-contaminated soils are one of the environmental problems that are thought to pose a major risk to the health of humans and other living things<sup>[40]</sup>.

Heavy metal pollution is one of the most serious environmental issues of all types of pollution. Arsenic (As), cadmium (Cd), chromium (Cr), copper (Cu), lead (Pb), mercury (Hg), nickel (Ni), and zinc (Zn) are the main heavy metal pollutants found in the environment<sup>[41, 42]</sup>.

One of the main things influencing the amount of heavy metals in the soil is the use of fertilisers. The addition of mineral additives to animal feed is another factor. Livestock requires minerals like Cu, Zn, Fe, Cr, Mn, and Co. Livestock are exposed to non-essential trace elements like cadmium,

mercury, arsenic, lead, and other heavy metals because mineral additives used in animal feed have low purity. The majority of heavy metals in feed are excreted by animals and used as "farmyard manure," which modifies the concentration of heavy metals in soil used on farms. Only trace amounts of these metals are absorbed by animals [60]. Earthworms in the soil absorb these heavy metals.

The important factors influencing the resistance phenotype in urban environments has been identified as metal pollution. The larvae pay a price in fitness when they have to adapt to the presence of metal pollutants in the environment. Under laboratory conditions, *An. gambiae* can be selected for metal tolerance quite quickly [43]. Nevertheless, there are substantial biological costs associated with this adaptation, such as decreased egg viability, immature survivorship and emergence, and decreased reproductive capacity [44].

#### 4.1 *In silico* docking of Metal – Protein Docking

In this docking study, HDOCK server was applied to dock the Metallothionein protein sequence of *Eisenia fetida* with Lead II sulfate. (Fig: 4). The application of HDOCK server to dock the lead II sulfate-containing Metallothionein protein sequence of *Eisenia fetida* is demonstrated in Figure 5. The server uses a hybrid algorithm that combines template-free and template-based docking to automatically predict the interaction between receptor and ligand molecules when data about them is entered. Unlike other similar docking servers, the HDOCK server can accept amino acid sequences as input. It also uses a hybrid docking strategy that allows experimental data about small-angle X-ray scattering and protein–protein binding sites to be included during the docking and post-docking phases [45, 46, 47, 48, 49, 50]. Table 1 shows the docking score between PbSO<sub>4</sub> and Metallothionein of *Eisenia fetida* which is -35.39 kcal/mol. Table 1 shows the docking score of -41.39 kcal/mol between PbSO<sub>4</sub> and Metallothionein of *An. gambiae*.

Figures 4, 5 clearly illustrates the molecular dynamics of how Metallothionein of *Eisenia fetida* binds with Lead II sulfate at various interacting sites. Fig.6,7,8 clearly represents the hydrophobic and electrostatic interaction between the metal and the protein of *Eisenia fetida*, viewed using Discovery Studio software. Figures: 11, 12 clearly illustrates the molecular dynamics of how Metallothionein of *An. gambiae* binds with Lead II sulfate at various interacting sites. Using Discovery Studio software, Fig: 13 depicts the hydrophobic and electrostatic interaction between the metal and the *An. gambiae*'s MT protein. The results of previous docking studies conducted in literature coincide with our findings [51, 52, 53, 54, 55, 56].

The results clearly show that the amino acids, (ALA:44,ASP:45,ALA:46,GLY:49,ASN:53,CYS:62,CYC:62,CYS:64,CYS:64,SER:65,GLY:67,SER:65,GLY:67,SER:68,CYS:69), are in charge of lead II sulphate (PbSO<sub>4</sub>) build-up inside the earthworm's body. 9-78 CYS\_RICH (PS50311): Drug binding Domain: 72-77 (GLY: 77) and 52-57 (ASN: 53). The results clearly show that the amino acids, CYS:65,CYS:6,CYS:6,CYS:10,CYS:12,CYS:12,CYS:27,CYS:27, are in charge of lead II sulphate (PbSO<sub>4</sub>) build-up inside the earthworm's body. (PROSITE: PS51257: Prokaryotic membrane lipoprotein lipid attachment site profile.) 1-23 is the motif range. The drug binding sites are present in the motif regions of MT of *An. gambiae*.

Previous research in the literature has demonstrated that the high cysteine residue content of the Metallothioneins found in

*Eisenia fetida* allows them to bind with a variety of heavy metals, most notably Lead II sulphate [57,58]. Furthermore, it has been demonstrated in wet lab studies that *Eisenia fetida*'s accumulation of lead II sulfate causes the protein Metallothionein to be expressed [59].

These findings are consistent with our ongoing *in silico* study, which has also demonstrated how the accumulation of the heavy metal, Lead II sulphate in *Eisenia fetida* and *An. gambiae* takes place and how it is expressed through the protein, Metallothionein.

#### 5. Conclusion

Our current *In silico* research investigation primarily focuses on the molecular interactions between lead II sulphate and the *Eisenia fetida* protein and the larvae of *An.gambiae* using state-of-the-art *In silico* study protocols. The overall findings of our study indicate that the expression of the Metallothionein protein of *Eisenia fetida* and larval *An.gambiae* is based on the accumulation of lead II sulphate metal. Ultimately, our current research has revealed that the binding of the metal Lead II sulphate with the two species results in adverse effects on both species, but in the case of mosquitoes, it benefits the human population, and in the case of *An. gambiae* larvae, it has an adverse effect on agriculture.

#### 6. Acknowledgement

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#### 7. References

1. Spurgeon DJ, Lawlor A, Hooper HL, Wadsworth R, Svendsen C, Thomas LD, *et al.* Outdoor and indoor cadmium distributions near an abandoned smelting works and their relations to human exposure. *Environ. Pollut.* 2011;159:3425-3432.
2. Ali H, Khan E, Sajad MA. Phytoremediation of heavy metals-Concepts and applications. *Chemosphere.* 2013;91:869-881.
3. Duffus JH. Heavy metals a meaningless term? (IUPAC Technical Report) *Pure Appl. Chem.* 2002;74:793-807.
4. Kemp DD. *The Environment Dictionary.* Psychology Press; Hove, UK; c1998.
5. Oves M, Khan MS, Zaidi A, Ahmad E. Toxicity of Heavy Metals to Legumes and Bioremediation. Springer; Berlin/Heidelberg, Germany. Soil contamination, nutritive value, and human health risk assessment of heavy metals: An overview; c2012. p. 1-27.
6. Li C, Zhou K, Qin W, Tian C, Qi M, Yan X, *et al.* A review on heavy metals contamination in soil: Effects, sources, and remediation techniques. *Soil Sediment Contam. Int. J.* 2019;28:380-394.
7. Chen H, Teng Y, Lu S, Wang Y, Wang J. Contamination features and health risk of soil heavy metals in China. *Sci. Total Environ.* 2015;512:143-153.
8. Li Z, Ma Z, van der Kuip TJ, Yuan Z, Huang L. A review of soil heavy metal pollution from mines in China: Pollution and health risk assessment. *Sci. Total Environ.* 2014;468:843-853.
9. Wu H, Yang F, Li H, Li Q, Zhang F, Ba Y, *et al.* Heavy metal pollution and health risk assessment of agricultural soil near a smelter in an industrial city in China. *Int. J.*

- Environ. Health Res. 2020;30:174-186.
10. Guo X. Cost of pollution in China: Economic estimates of physical damages. World Bank. 2007;39236:1-151.
  11. Hou D, O'Connor D, Nathanail P, Tian L, Ma Y. Integrated GIS and multivariate statistical analysis for regional scale assessment of heavy metal soil contamination: A critical review. Environ. Pollut. 2017;231:1188-1200.
  12. Solgi E, Esmaili-Sari A, Riyahi-Bakhtiari A, Hadipour M. Soil contamination of metals in the three industrial estates, Arak, Iran. Bull. Environ. Contam. Toxicol. 2012;88:634-638.
  13. Burges A, Epelde L, Garbisu C. Impact of repeated single-metal and multi-metal pollution events on soil quality. Chemosphere. 2015;120:8-15.
  14. Zhang P, Qin C, Hong X, Kang G, Qin M, Yang D, *et al.* Risk assessment and source analysis of soil heavy metal pollution from lower reaches of Yellow River irrigation in China. Sci. Total Environ. 2018;633:1136-1147.
  15. Giller K, McGrath S. Pollution by toxic metals on agricultural soils. Nature. 1988;335:676.
  16. Rodrigues S, Cruz N, Coelho C, Henriques B, Carvalho L, Duarte A, *et al.* Risk assessment for Cd, Cu, Pb and Zn in urban soils: Chemical availability as the central concept. Environ. Pollut. 2013;183:234-242.
  17. Zhou J, Liang J, Hu Y, Zhang W, Liu H, You L, *et al.* Exposure risk of local residents to copper near the largest flash copper smelter in China. Sci. Total Environ. 2018;630:453-461. DOI: 10.1016/j.scitotenv.2018.02.211.
  18. Shi T, Ma J, Zhang Y, Liu C, Hu Y, Gong Y, *et al.* Status of lead accumulation in agricultural soils across China (1979–2016) Environ. Int. 2019;129:35-41.
  19. Zhao FJ, Ma Y, Zhu YG, Tang Z, McGrath SP. Soil contamination in China: Current status and mitigation strategies. Environ. Sci. Technol. 2015;49:750-759. DOI: 10.1021/es5047099.
  20. Adnan M, Xiao B, Xiao P, Zhao P, Bibi S. Heavy Metal, Waste, COVID-19, and Rapid Industrialization in This Modern Era—Fit for Sustainable Future. Sustainability. 2022;14:4746.
  21. Mielke HW, Reagan PL. Soil is an important pathway of human lead exposure. Environ. Health Perspect. 1998;106(Suppl. S1):217-229.
  22. Wang S, Zhang J. Blood lead levels in children, China. Environ. Res. 2006;101:412-418.
  23. Qiu K, Xing W, Scheckel K, Cheng Y, Zhao Z, Ruan X, *et al.* Temporal and seasonal variations of As, Cd and Pb atmospheric deposition flux in the vicinity of lead smelters in Jiuyan, China. Atmos. Pollut. Res. 2016;7:170-179.
  24. Zirbes L, Brostaux Y, Mescher M, Jason M, Haubruge E, Deneubourg JL, *et al.* Self-Assemblage and quorum in the earthworm *Eisenia fetida* (Oligochaete, Lumbricidae). PLOS ONE. 2012;7(3):e32564. DOI: 10.1371/journal.pone.0032564.
  25. Kumar JN, Soni H, Kumar RN, Patil N. Growth and reproduction of *Eisenia foetida* in various industry waste sludge during vermicomposting: a laboratory investigation. Int J Environ Waste Manag. 2010;5(3-4):379-91.
  26. Sinka M, Bangs M, Manguin S, Coetzee M, Mbogo C, Hemingway J, *et al.* The dominant Anopheles vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic precis. Parasit Vectors. 2010;3(1):117
  27. Awolola TS, Oduola AO, Obansa JB, Chukwura NJ, Unyimadu JP. Anopheles gambiae s. s. breeding in polluted water bodies in urban Lagos, southwestern Nigeria. J Vector Borne Dis. 2007;44(4):241-244.
  28. Djouaka RF, Bakare AA, Bankole HS, Doannio JM, Kossou H, Akogbeto MC, *et al.* Quantification of the efficiency of treatment of Anopheles gambiae breeding sites with petroleum products by local communities in areas of insecticide resistance in the Republic of Benin. Malar J. 2007;6:56. Published 2007 May 8. doi:10.1186/1475-2875-6-56
  29. Mireji PO, Keating J, Kenya E. Differential Induction of Proteins in Anopheles gambiae sensu stricto (Diptera: Culicidae) Larvae in Response to Heavy Metal Selection. Int J Trop Insect Sci. 2006;26(4):214-226. DOI:10.1017/S1742758406658955
  30. Antonio-Nkondjio C, Fossog BT, Ndo C. Anopheles gambiae distribution and insecticide resistance in the cities of Douala and Yaoundé (Cameroon): influence of urban agriculture and pollution. Malar J. 2011;10:154. Published 2011 Jun 8. doi:10.1186/1475-2875-10-154
  31. Jones CM, Toé HK, Sanou A. Additional selection for insecticide resistance in urban malaria vectors: DDT resistance in Anopheles arabiensis from Bobo-Dioulasso, Burkina Faso. PLoS One. 2012;7(9):e45995. DOI:10.1371/journal.pone.0045995
  32. Poupardin R, Reynaud S, Strode C, Ranson H, Vontas J, David JP, *et al.* Cross-induction of detoxification genes by environmental xenobiotics and insecticides in the mosquito Aedes aegypti: impact on larval tolerance to chemical insecticides. Insect Biochem Mol Biol. 2008;38(5):540-551. doi:10.1016/j.ibmb.2008.01.004
  33. Poupardin R, Riaz MA, Jones CM, Chandor-Proust A, Reynaud S, David JP, *et al.* Do pollutants affect insecticide-driven gene selection in mosquitoes? Experimental evidence from transcriptomics. Aquat Toxicol. 2012;114-115:49-57. DOI:10.1016/j.aquatox.2012.02.001
  34. Riaz MA, Poupardin R, Reynaud S, Strode C, Ranson H, David JP, *et al.* Impact of glyphosate and benzo[a]pyrene on the tolerance of mosquito larvae to chemical insecticides. Role of detoxification genes in response to xenobiotics. Aquat Toxicol. 2009;93(1):61-69. doi:10.1016/j.aquatox.2009.03.005
  35. Nkya TE, Poupardin R, Laporte F. Impact of agriculture on the selection of insecticide resistance in the malaria vector Anopheles gambiae: a multigenerational study in controlled conditions. Parasit Vectors. 2014;7:480. Published 2014 Oct 16. doi:10.1186/s13071-014-0480-z
  36. Schoch CL, Ciuffo S, Domrachev M. NCBI Taxonomy: a comprehensive update on curation, resources and tools. Database (Oxford). 2020;2020:baaa062.
  37. Yuvaraj A, Govarathanan M, Karmegam N. Metallothionein dependent-detoxification of heavy metals in the agricultural field soil of industrial area: Earthworm as field experimental model system [published correction appears in Chemosphere. 2021 Jun;273:130289]. Chemosphere. 2021;267:129240.
  38. Barassi GM, Klimsa M, Borrmann T, Cairns MJ, Kinkel J, Valenzuela F, *et al.* Lead sulfate nano- and microparticles in the acid plant blow-down generated at the sulfuric acid

- plant of the El Teniente mine, Chile. *Environ Sci Process Impacts*. 2014;16(12):2734-2741.
39. Yan Y, Tao H, He J, Huang S-Y. The HDOCK server for integrated protein-protein docking. *Nature Protocols*; c2020.
  40. Parihar, Kapil, Kumar, Rajeev, Sankhla, Singh M, *et al.* Impact of Heavy Metals on Survivability of Earthworms (November 26, 2019). *International Medico-Legal Reporter Journal*. 2019, 2(3). Available at SSRN: <https://ssrn.com/abstract=3497689>
  41. Yang YT, Chen Y, Leng F, Huang L, Wang ZJ, Tian WQ, *et al.* Recent Advances on Surface Modification of Halloysite Nanotubes for Multifunctional Applications. *Appl. Sci*. 2017, 7(12).
  42. Khan MA, Amir RM, Ahmad A. Application of nanoparticles for the removal of heavy metals from wastewater. *Int. J. Agric. Food Sci*. 2022;4(2):109-113. DOI: 10.33545/2664844X.2022.v4.i2b.102
  43. Mireji PO, Keating J, Kenya E. Differential Induction of Proteins in *Anopheles gambiae sensu stricto* (Diptera: Culicidae) Larvae in Response to Heavy Metal Selection. *Int J Trop Insect Sci*. 2006;26(4):214-226. DOI:10.1017/S1742758406658955
  44. Mireji PO, Keating J, Hassanali A. Biological cost of tolerance to heavy metals in the mosquito *Anopheles gambiae*. *Med Vet Entomol*. 2010;24(2):101-107. doi:10.1111/j.1365-2915.2010.00863.x
  45. Remmert M, Biegert A, Hauser A, Soing J. HHblits: lightning-fast iterative protein sequence searching by HMM-HMM alignment. *Nat Methods*. 2011;9:173-175.
  46. Pearson WR, Lipman DJ. Improved tools for biological sequence comparison. *Proc Natl Acad Sci USA*. 1988;85:2444-2448.
  47. Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, *et al.* Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Mol Syst Biol*. 2011;7:539.
  48. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, *et al.* Clustal W and Clustal X version 2.0. *Bioinformatics*. 2007;23:2947-8.
  49. Marti-Renom MA, Stuart A, Fiser A, Sanchez R, Melo F, Sali A, *et al.* Comparative protein structure modeling of genes and genomes. *Annu Rev Biophys Biomol Struct*. 2000;29:291-325.
  50. Berman HM, Westbrook J, Feng Z, Gilliland G, Bhat TN, Weissig H, *et al.* The Protein Data Bank. *Nucleic Acids Res* 2000; 28:235-242.
  51. Astalakshmi P, John MB, Kavinilavu G, Vimala D. Identification of the efficiency of Pentane on the bacterial and insecticide proteins of *Aedes aegypti* and *Aeromonas hydrophila* by In silico methods. *Int J Mosq Res*. 2023;10(3):47-53. DOI: <https://doi.org/10.22271/23487941.2023.v10.i3a.6782>.
  52. Astalakshmi P, John MB, Kavinilavu G, Vimala D. In silico study on Hexadecanoic acid against the outer membrane protein transport protein of *Culex quinquefasciatus* and *Aeromonas hydrophila*. *Int J Mosq Res*. 2023;10(4):07-14. DOI:<https://doi.org/10.22271/23487941.2023.v10.i4a.68>.
  53. Nijanthi P, Santhi S, Munivelan B. Molecular dynamics studies on the arginine kinase protein of *Aedes sollicitans*: Against the natural chemical compound, Gedunin. *Int J Mosq Res*. 2023;10(2):10-14.
  54. Maithreyee S1, Prabha V1, Molecular Interactions between Anti-Inflammatory Drug with Colorectal Cancer (MSH2) Protein Using In-silico Studies. *Solovyov Studies ISPU*. 2023, 71(10). DOI:10.37896/ispu71.10/012.
  55. Nithya G, Prabha V. Identification of a plant derivative (*Hibiscus cannabinus*) for mosquito (*Anopheles darlingi*) control using in silico protein-protein docking techniques. *Int J Mosq Res*. 2023;10(4):25-29. DOI:
  56. Munivelan B, Identification of the mutated sites present in the transmembrane regions of SCN1A\_HUMAN (Sodium Voltage-Gated Channel Alpha Subunit 1) using In silico techniques. *Int. J Pharm. Int. Biosci*. 2020;5(1):1-6.
  57. Gruber C, Stürzenbaum S, Gehrig P. Isolation and characterization of a self-sufficient one-domain protein. (Cd)-Metallothionein from *Eisenia foetida*. *European Journal of Biochemistry*. 2000 Jan;267(2):573-582.
  58. Brulle F, Mitta G, Cocquerelle C. Cloning and real-time PCR testing of 14 potential biomarkers in *Eisenia fetida* following cadmium exposure. *Environmental Science & Technology*. 2006 Apr;40(8):2844-2850.
  59. Mostafaii G, Aseman E, Asgharnia H, Akbari H, Iranshahi L, Sayyaf H, *et al.* Efficiency of the earthworm *Eisenia fetida* under the effect of organic matter for bioremediation of soils contaminated with cadmium and chromium. *Brazilian journal of chemical engineering*. 2016;33(4):827-834.
  60. Feng Z, Zhu H, Deng Q, He Y, Li J, Yin J, *et al.* Environmental pollution induced by heavy metal (loid) s from pig farming. *Environ. Earth Sci*. 2018, 77(3).