

ISSN: 2348-5906

CODEN: IJMRK2

IJMR 2023; 10(4): 43-53

© 2023 IJMR

<https://www.dipterajournal.com>

Received: 07-05-2023

Accepted: 18-06-2023

Baindur SiddharthFaculty of Medicine in Pilsen,
Charles University, Czech
Republic**Gohil Rinkesh**Kishinchand Chellaram College
of Science, Arts and Commerce,
Mumbai, Maharashtra, India**Kolte Satish**Kishinchand Chellaram College
of Science, Arts and Commerce,
Mumbai, Maharashtra, India**Corresponding Author:****Kolte Satish**Kishinchand Chellaram College
of Science, Arts and Commerce,
Mumbai, Maharashtra, India

International Journal of Mosquito Research

Mosquito repellence activity of solvent extracts and activated charcoal obtained from agro waste- coconut shells

Baindur Siddharth, Gohil Rinkesh and Kolte Satish

DOI: <https://doi.org/10.22271/23487941.2023.v10.i4a.686>

Abstract

Burning Coconut shells have been used since ages in rural India to ward away mosquitoes; we study this traditional method in detail in our research, using modern techniques such as dry distillation, solvent extraction, chromatography and infrared spectroscopy to decipher its constitutional contents and efficacy scientifically. In our research, coconut shells in the form of activated charcoal and chemical extracts (solvent extracts of petroleum ether, chloroform and methanol) obtained from the shell itself were used to test against their mosquito repellence and knock off efficacy in a Peet Grady's chamber as per WHO guidelines to test insecticides. The petroleum ether extract was found to be most efficacious with 100% mortality rate in 30 mins. The extract mainly contained aromatic compounds, alkanes and ketonic groups deduced using the abovementioned chemical means.

Keywords: Solvent extracts, charcoal obtained, coconut shells

Introduction

Mosquitoes are considered as a pest to humankind in all parts of the world. They serve as a major vector for spreading diseases, not only through pathogens but also through parasites [2]. Mosquitoes are responsible for the propagation of diseases, specifically malaria, chikungunya, dengue, Japanese encephalitis, West Nile, yellow fever, and Zika throughout the world [3]. Around 400,000 deaths are caused per year due to malaria [2]. Even though constant developments at generating vaccines for mosquito-borne arboviral diseases are in way, either as inactivated vaccines, viral-vector vaccines, live attenuated vaccines, protein vaccines, or nucleic acid vaccines, approximately half of the world's population is expected to be at the risk of arbovirus transmission by 2050 [4]. This raises an alarming note for the need for pest control and eradication or at the least control of its propagating vector i.e., mosquitoes.

Series of literature suggest the use of natural ingredients (including *Zanthoxylum limonella*, *Azadirachta indica*, *Tinospora rumphii*, *Citrus grandis*, *Jatropha curcas*, *Cymbopogon nardus*, and *Cocos nucifera*) used to repel or eradicate mosquito larvae and adults [2, 5-9].

In rural India, coconut (*Cocos nucifera*) coir or shell fibres have been used in bonfires or small flames to ward away mosquitoes; a few select references suggest the use of coconut derived compounds as efficient mosquito repellents as well [5-12]. This was the very hypothesis for our research i.e. to find any active component in the traditional ways of repelling mosquitoes and whether it can be used for commercial purposes.

The coconut tree is affectionately called the *kalpavriksha*- the mythical wish fulfilling tree, by coastal Indians, as each part of the tree can be used for various purposes. Besides the fruit having nutritious value, research has also shown the use of coconut fibres, kernels, leaves and bark in production, purification, and isolation of various chemicals: it has been used as an industrial filler, in water purification to remove bacteria, organic pollutants, and heavy metals and even in the generation of nanoparticles like magnetite and palladium [12-19]. Based on our research and past literature, we assess the use of coconut shell and its isolated extracts in its efficiency as active ingredients, by part, or as whole, in mosquito repellent and mosquito-cidal activity.

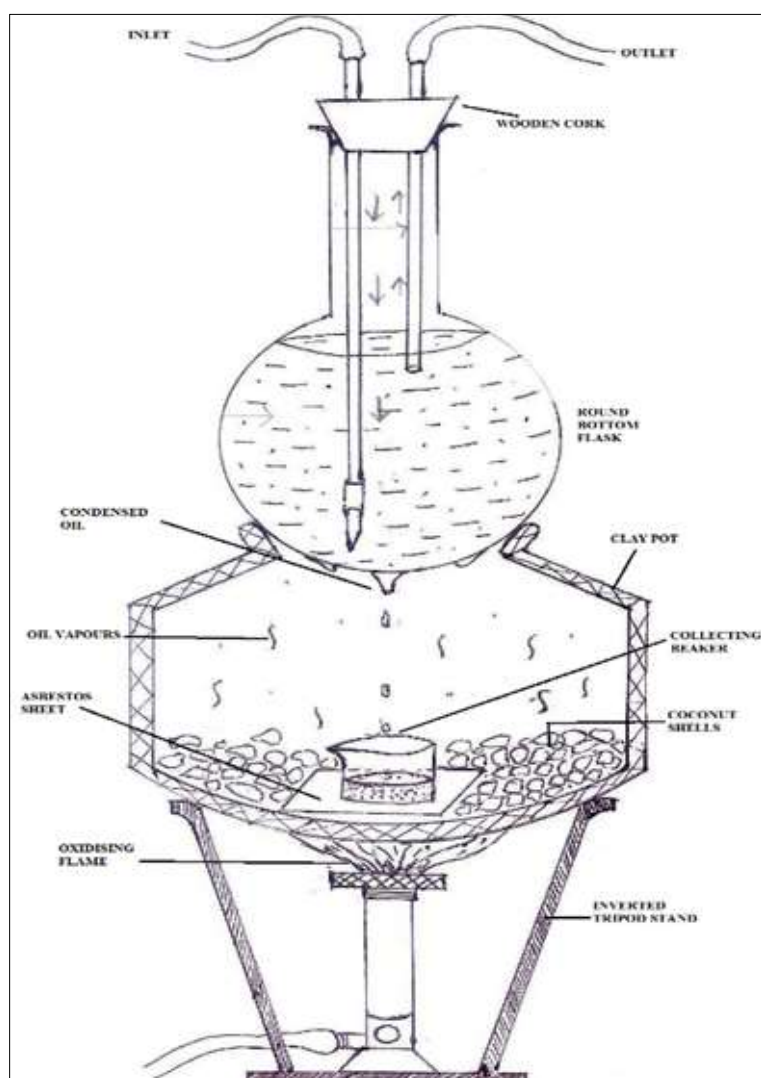


Fig 1: Fractional dry distillation of coconut shells to obtain crude coconut shell oil which is further processed by solvent extraction to obtain the solvents E1, E2 and E3.

Materials & Methods

Preparation of coconut shells extract and charcoal

Around 1kg of *Cocos nucifera* shells were sourced from local vendors in Mumbai district. They were peeled, dried and broken into large pieces of about 3 x 3 cm each. The pieces were roasted gradually in an earthen pot apparatus as described in fig 1, for 8hrs at high temperatures to remove moisture and organic oils. The organic oils were collected as (E) which would be processed and characterised through solvent extraction and other processes. The shell residue obtained was crushed into fine particles using mortar and pestle. The particles were filtered through a metal sieve to obtain very fine and uniform particle size (diameter of around 1mm).

Activation of charcoal

The charcoal obtained from the above process and wood charcoal (sourced from local vendors) crushed into fine uniform powder was soaked in methanol overnight to separate organic impurities. It was then filtered, dried, and heated at 1000 C in an oven for 3hrs. This clears off volatile substances and leaves behind pure charcoal carbon.

Solvent extraction

The crude oil obtained from the dry distillation of the shells

(yield of 1 kg of coconut shells gives about 250cc of crude extract). This crude oil obtained was split for characterization and experimentation. 25cc of oil was separated by solvent extraction using three solvents with increasing polarities i.e. petroleum ether (non-polar), chloroform (polar) and methanol (highly polar). Different extracts obtained using these three solvents were collected and air dried to evaporate excess solvent. After complete drying, each extract was named E₁, E₂ and E₃ respectively.

Fractional distillation

A part of the crude oil was distilled on a sand bath using a water condenser to obtain distillates D1-D4 at temperatures 60C, 80C, 90C and 100C respectively. These distillates were then characterized further by Fourier Transform Infrared (FTIR) spectroscopy and silica gel-thin layered chromatography (TLC) to study the level of purity. (Supplementary data: D1-D4).

Column chromatography

A part of the dried petroleum ether extract E₁ (which showed maximum mortality in the mortality tests) was further fractionated using silica gel column chromatography using gradual ratios of non-polar- polar solvents Petroleum ether-Chloroform-Methanol to obtain 6 fractions, which were

further characterized by Fourier Transform infrared (FTIR) spectroscopy and its purity was checked using silica gel-thin layered chromatography (TLC). (Supplementary data: A-F).

Preparation of mosquito repellent sticks

To study the correlation between mosquito repellent activity, Jigat (*Machilus macrantha*: inert adhesive filler) powder, saw dust and coconut or wood charcoal powder were mixed in a ratio of 1:1:1 by weight and kneaded with water to obtain dough. This dough was then rolled onto bamboo sticks (average weight of each stick = 0.5g) to obtain sticks of uniform length, breadth and weight. They were then air dried for 24 hrs. The obtained sticks were of average length 11 cm, breadth 0.5 cm and weight 1.5g each.

36 sticks with coconut charcoal and 36 sticks with wood charcoal were prepared

The dried extracts E1- E2 and E3 obtained after solvent extraction were dissolved using methanol and the prepared sticks were dipped in their respective solutions and air-dried to evaporate the methanol. The sticks are now impregnated with extracts E1, E2 and E3. The composition is as follows:

| Extract | Wood charcoal (WC) | Coconut Charcoal (CC) |
|---------|--------------------|-----------------------|
| E1 | 12 sticks | 12 sticks |
| E2 | 12 sticks | 12 sticks |
| E3 | 12 sticks | 12 sticks |

Mosquito repellence and mortality test: The mosquito repellent tests were performed at the Haffkine Institute, Mumbai under the WHO guidelines for testing the efficacy of insecticide products on mosquitoes [1]. Tests were performed on 50 non-blood-fed, 2–5-day old female *Aedes aegypti* species of mosquitoes in a Peet Grady chamber of internal volume 180cc, at 27 C ± 2 C, 80% ± 10% relative humidity (RH) and photoperiod 12:12 h (Light: dark). The repellent sticks were lit after the mosquitoes are kept captivated in the chamber, at T=0 and the tests were carried out for a total of 30 mins from being incensed. Knockdown readings were recorded for every 5 min up to T=30min and mortality of mosquitoes was calculated by the formula:

$$\text{Mortality (\%)} = \frac{\text{No. of mosquitoes knocked out (for 24hr)}}{\text{Total mosquitoes in chamber}} \times 100$$

Results and Discussion

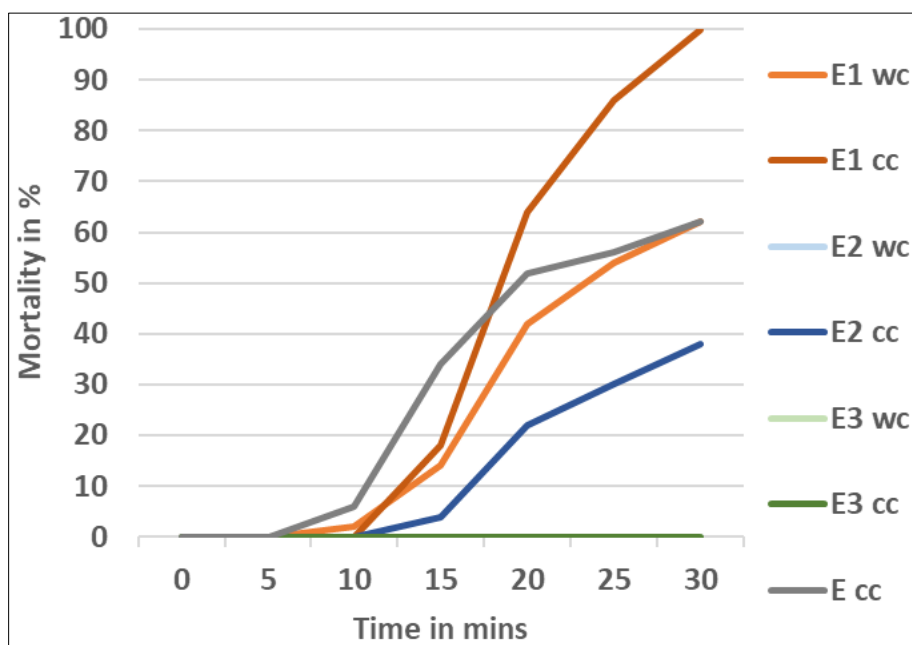


Fig 2: Mortality rates of the solvent extracts (E1 in Petroleum Ether, E2 in Chloroform and E3 in Methanol and E is crude coconut shell oil; (diluent used is Methanol) in wood charcoal (wc) or coconut charcoal (cc) sticks

As observed in Fig. 2, the mortality rates were highest in the (E1 cc) coconut charcoal sticks dipped in petroleum ether extracts compared to its wood charcoal counterparts. Coconut charcoal enhanced the ability to repel mosquitoes much more efficiently than the wood charcoal filled repellent sticks. The repellence activity between the three solvent extracts was in the order of Petroleum Ether > Chloroform > Methanol, with methanol having the least activity. Crude oil extract (E) with coconut charcoal filler, showed average mosquito repellence. Isolated Petroleum Ether (E1) extract showed repellence higher than crude extract and thus was more refined or free of any inhibitory substances. The characterization of the E1 extract showed presence of alkanes, ketonic groups and aromatic hydrocarbons (supplementary data). The fractions

isolated from fractional distillation and column chromatography could be further tested against its mosquito repellence activities in future prospects.

References

1. Indd A. Guidelines for testing the efficacy of insecticide products used in aircraft Control of Neglected Tropical Diseases WHO Pesticide Evaluation Scheme and Department of Global Capacity, Alert and Response Support for International Health Regulations Capacity Development Ports, airports and ground crossing; c2012.
2. Jones RT, Ant TH, Cameron MM, Logan JG. Novel control strategies for mosquito-borne diseases: Control of mosquito-borne diseases. Philosophical Transactions of

- the Royal Society B: Biological Sciences, 2021, 376 Preprint at <https://doi.org/10.1098/rstb.2019.0802>.
- Lee H, Halverson S, Ezinwa N. Mosquito-Borne Diseases. Primary Care: Clinics in Office Practice. 2018;45:393-407.
 - Kraemer MUG, *et al.* Past and future spread of the arbovirus vectors *Aedes aegypti* and *Aedes albopictus*. Nat Microbiol. 2019;4:854-863.
 - Zhang S, *et al.* Larvicidal activity of natural repellents against the dengue vector, *Aedes aegypti*. <http://meridian.allenpress.com/jamca/article-pdf/36/4/227/2774457/i8756-971x-36-4-227.pdf>.
 - Pohlit AM, Lopes NP, Gama RA, Tadei WP, De Andrade Neto VF. Patent literature on mosquito repellent inventions which contain plant essential oils - A review. Planta Medica. 2011;77:598-617 Preprint at <https://doi.org/10.1055/s-0030-1270723>.
 - Das NG, Baruah I, Talukdar PK, Das SC. Evaluation of botanicals as repellents against mosquitoes. J Vect Borne Dis vol. 40.
 - Innocent E, *et al.* Anti-mosquito plants as an alternative or incremental method for malaria vector control among rural communities of Bagamoyo District, Tanzania. J Ethnobiol Ethnomed. 2014, 10.
 - Gutierrez PM, Antepuesto AN, Eugenio AL, Fleurelle M, Santos L. Larvicidal Activity of Selected Plant Extracts against the Dengue vector *Aedes aegypti* Mosquito. International Research Journal of Biological Sciences, 2014, 3 www.isca.me.
 - Zhu JJ, *et al.* Better than DEET Repellent Compounds Derived from Coconut Oil. Sci Rep. 2018, 8.
 - Zhang L, *et al.* Using charcoal as base material reduces mosquito coil emissions of toxins. Indoor Air. 2010;20:176-184.
 - Elango G, *et al.* Cocos nucifera coir-mediated green synthesis of Pd NPs and its investigation against larvae and agricultural pest. Artif Cells Nanomed Biotechnol. 2017;45:1581-1587.
 - Mrozik W, *et al.* Valorisation of agricultural waste derived biochars in aquaculture to remove organic micropollutants from water - experimental study and molecular dynamics simulations. J Environ Manage, 2021, 300.
 - Sebastian A, Nangia A, Prasad MNV. A green synthetic route to phenolics fabricated magnetite nanoparticles from coconut husk extract: Implications to treat metal contaminated water and heavy metal stress in *Oryza sativa* L. J Clean Prod. 2018;174:355-366.
 - Bhatnagar A, Vilar VJP, Botelho CMS, Boaventura RAR. Coconut-based biosorbents for water treatment-A review of the recent literature. Advances in Colloid and Interface Science. 2010;160:1-15 Preprint at <https://doi.org/10.1016/j.cis.2010.06.011>.
 - Goel J, Kadirvelu K, Rajagopal C Mercury. (II) removal from water by coconut shell based activated carbon: Batch and column studies. Environmental Technology (United Kingdom). 2004;25:141-153.
 - Van Der Mei HC, *et al.* Influence of adhesion to activated carbon particles on the viability of waterborne pathogenic bacteria under flow. Biotechnol Bioeng. 2008;100:810-813.
 - Wu Y, *et al.* Comparative and competitive adsorption of Cr (VI), As (III), and Ni (II) onto coconut charcoal. Environmental Science and Pollution Research. 2013;20:2210-2219.
 - Tan IAW, Ahmad AL, Hameed BH. Preparation of activated carbon from coconut husk: Optimization study on removal of 2,4,6-trichlorophenol using response surface methodology. J Hazard Mater. 2008;153:709-717.

Supplementary Data

Typical Ranges of Frequencies in Fourier Transform Infrared Spectroscopy

| Bond | Compound Type | Frequency range, cm ⁻¹ |
|-----------------|--|-----------------------------------|
| C-H | Alkanes | 2960-2850 |
| | | 1470-1350 |
| | CH ₂ Umbrella Deformation | 1380 |
| C-H | Alkenes | 3080-3020 |
| | | 1000-675 |
| C-H | Aromatic Rings | 3100-3000 |
| | Phenyl Ring Substitution Bands | 870-675 |
| | Phenyl Ring Substitution Overtones | 2000-1600 |
| C-H | Alkynes | 3333-3267 |
| | | 700-610 |
| C=C | Alkenes | 1680-1640 |
| C≡C | Alkynes | 2260-2100 |
| C=C | Aromatic Rings | 1600, 1500 |
| C-O | Alcohols, Ethers, Carboxylic acids, Esters | 1260-1000 |
| C=O | Aldehydes, Ketones, Carboxylic acids, Esters | 1760-1670 |
| O-H | Monomeric -- Alcohols, Phenols | 3640-3160 |
| | Hydrogen-bonded -- Alcohols, Phenols | 3600-3200 |
| | Carboxylic acids | 3000-2500 |
| N-H | Amines | 3500-3300 |
| | | 1650-1580 |
| C-N | Amines | 1340-1020 |
| C-N | Nitriles | 2260-2220 |
| NO ₂ | Nitro Compounds | 1660-1500 |
| | | 1390-1260 |

Chemical Analysis of each fraction**Fraction D₁****1. Organic spotting**

- Boiling range:** <60 °C
- Colour:** Light yellow
- Odour:** Pleasant
- Saturation test:** Saturated compound
- Elemental analysis:** C, H, (O)
- Determination of functional group:** Ketonic group

2. Chromatographic study

TLC: Single spot

Fraction D₂**Organic spotting**

- Boiling range:** 60-80 °C
- Colour:** Light brown
- Odour:** Pungent
- Saturation test:** Saturated compound
- Elemental analysis:** C, H, (O).
- Determination of functional group:** Carboxylic group present.

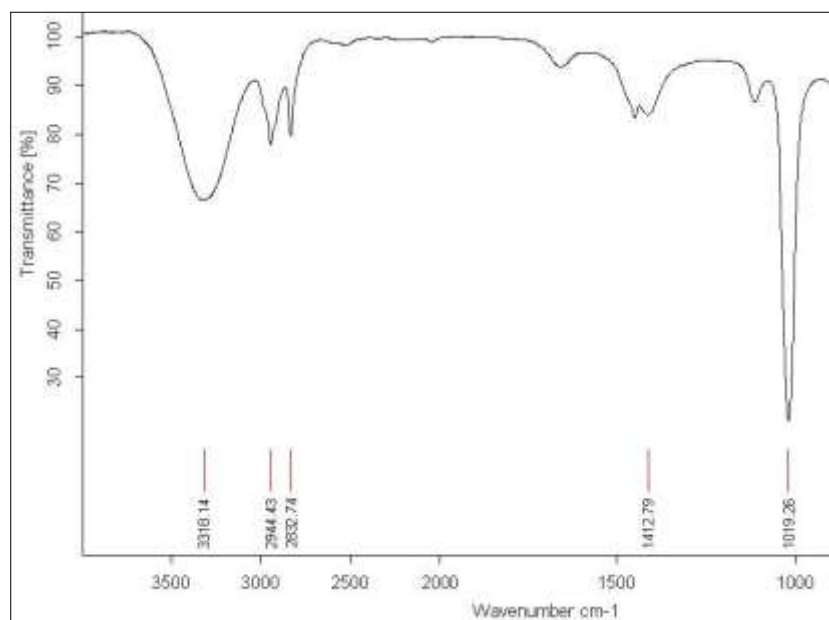
Chromatographic separation

TLC: shows two distinct spots. Therefore, further separation was done by column chromatography.

Chromatographic separation of B

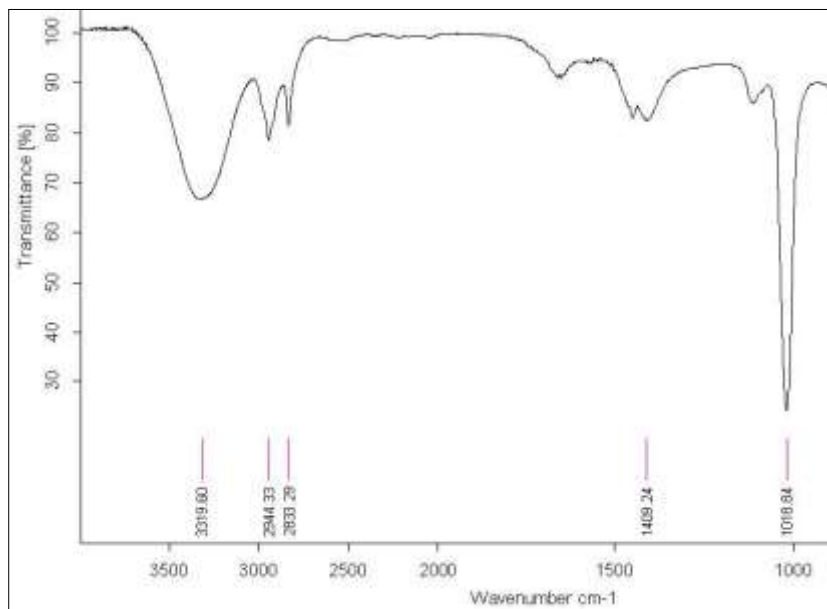
Fraction B was adsorbed on 1gm activated silica gel and loaded on a silica gel column of 30 cm in length. Various fractions collected and monitored using TLC techniques are showed in tabular form below.

| Test tube nos. | Composition of Mobile Phase | TLC results |
|-------------------|------------------------------|--------------------|
| 1-18 | Pet. Ether (100%) | - |
| 19-20 | Pet. Ether: Chloroform (9:1) | - |
| 21 | Pet. Ether: Chloroform (8:2) | - |
| 22 | Pet. Ether: Chloroform (7:3) | - |
| 23 | Pet. Ether: Chloroform (6:4) | - |
| 24-26 (Distilled) | Pet. Ether: Chloroform (5:5) | Single spot |
| 27 | Pet. Ether: Chloroform (3:7) | - |
| 28 | Chloroform: Methanol (7:3) | - |
| 29 | Methanol (100%) | Two distinct spots |

Fraction 24-26

| Sr. No. | Peaks | Functional | Group |
|---------|---------|-----------------|--------------------------|
| | | Structural Form | Names |
| 1. | 3318.14 | O-H | Alcohols |
| 2. | 2944.43 | C-H | Alkanes |
| 3. | 2832.74 | C-H | Alkanes |
| 4. | 1412.79 | C-H | Alkenes |
| 5. | 1019.26 | C-O | Ethers, Carboxylic Acids |

Fraction 29



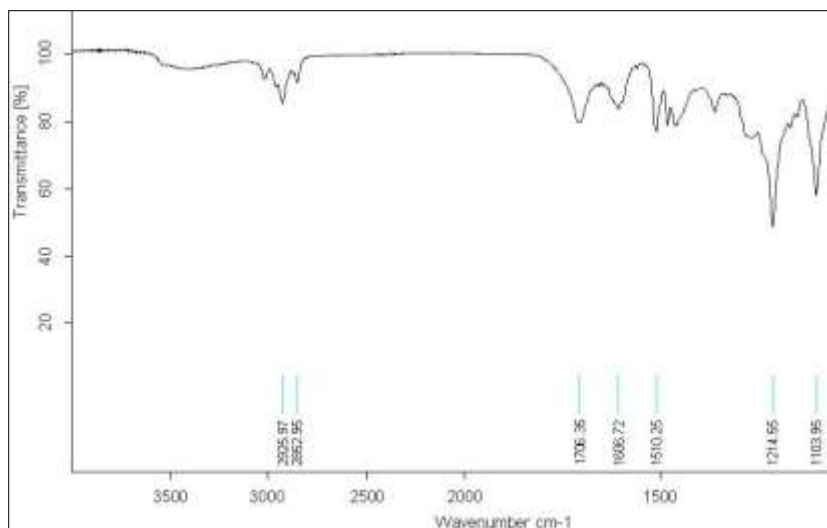
| Sr. No. | Peaks | Functional | Group |
|---------|---------|-----------------|--------------------------|
| | | Structural Form | Names |
| 1. | 3319.60 | O-H | Alcohols |
| 2. | 2944.33 | C-H | Alkanes |
| 3. | 2833.29 | C-H | Alkanes |
| 4. | 1409.24 | C-H | Alkenes |
| 5. | 1018.84 | C-O | Ethers, Carboxylic Acids |

Fraction D₃

1. Organic spotting

- Boiling range: 80-90 °C
- Colour: brown
- Odour: pungent
- Saturation test: saturated compound
- Elemental analysis: C, H, (O).
- Determination of functional (since the quantity remaining was not suitable for classical methods so further information about functional group was collected using IR spectroscopy)

2. Chromatographic separation



TLC: Two distinct spots. (Fraction size was too small so column chromatography was not implemented)

3. IR Spectrum

| Sr. No. | Peaks | Functional Structural Form | Group Names |
|---------|---------|----------------------------|---------------------------------|
| 1. | 2925.97 | C-H | Alkanes |
| 2. | 2852.95 | O-H | Carboxylic Acids |
| 3. | 1606.72 | C-H | Phenyl Ring Substitution |
| 4. | 1510.25 | NO ₂ | Nitro Compound |
| 5. | 1214.55 | C-O | Alcohol, Ester, Carboxylic Acid |
| 6. | 1103.95 | C-O | Alcohol, Ester, Carboxylic Acid |

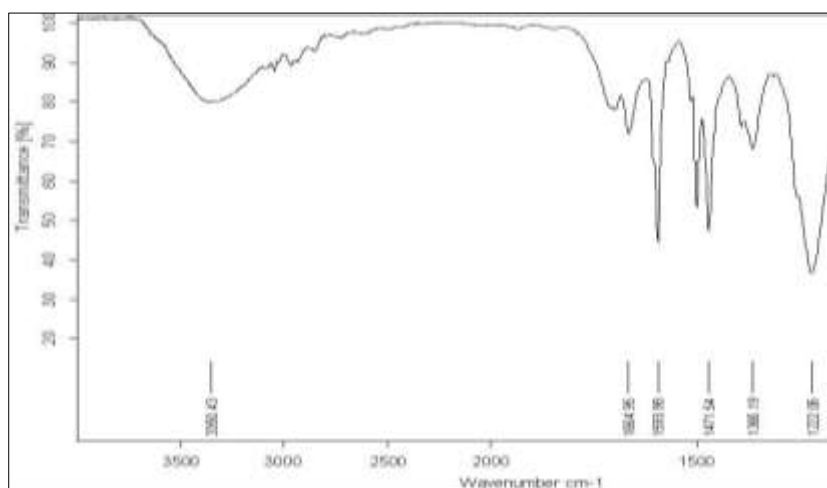
Fraction D₄

1. Organic spotting

- Colour: brown
- Odour: pungent
- Saturation test: saturated compound.
- Elemental analysis: C, H, (O).
- Determination of functional group: OH
- Boiling range: 90-98°C

2. Chromatographic separation

(Fraction size was too small so column chromatography was not implemented)



IR Spectrum

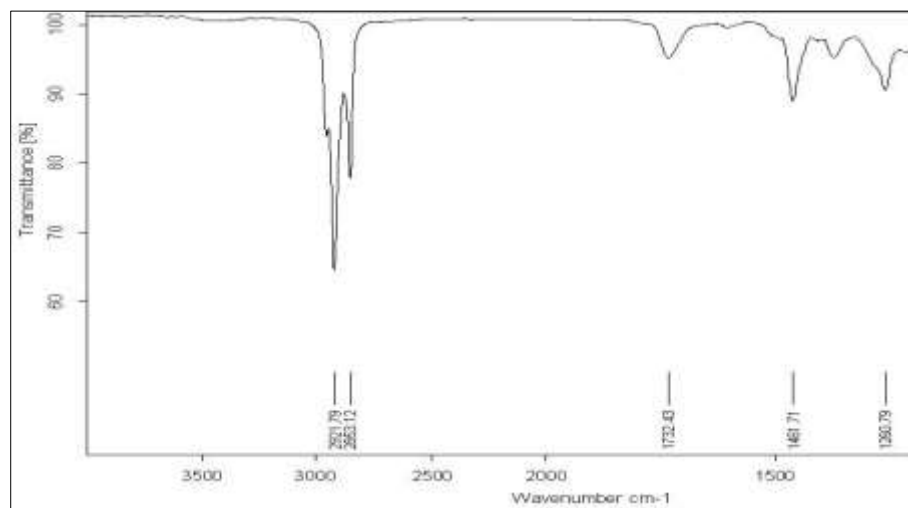
| Sr. No. | Peaks | Functional Group | Names |
|---------|---------|------------------|---------------------------------|
| 1. | 3350.43 | O-H | Monomeric - Alcohols, Phenols |
| 2. | 1664.95 | C=C | Alkenes |
| 3. | 1593.98 | N-H | Amines |
| 4. | 1471.54 | C=C | Aromatic Ring |
| 5. | 1366.19 | NO ₂ | Nitro Compounds |
| 6. | 1222.06 | C-O | Alcohol, Ester, Carboxylic Acid |

Silica gel Column Chromatography of Petroleum Ether Extract E1

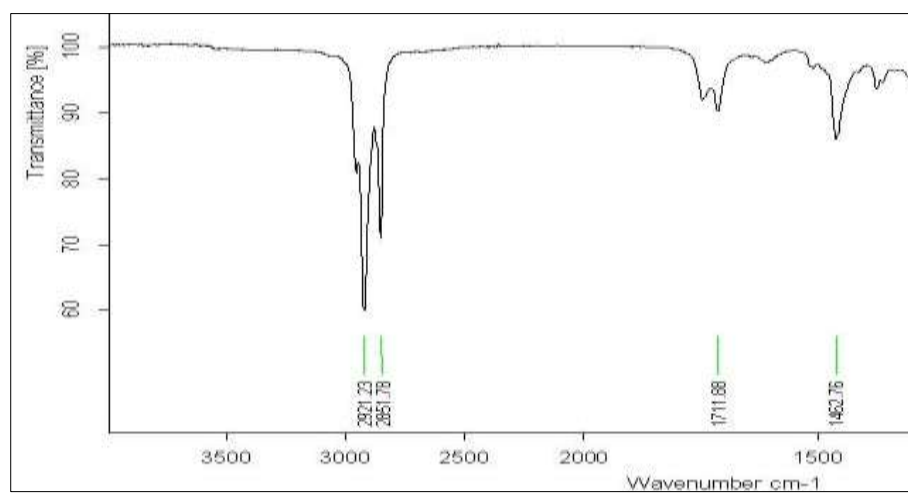
| Test tube no. | Composition of Mobile phase | TLC results | Fractions |
|-----------------------|--------------------------------|-------------|------------------|
| 1-5 | Pet. Ether (100%) | -- | -- |
| 6-17 | Pet. Ether: Chloroform (98:2) | -- | -- |
| 18-25 | Pet. Ether: Chloroform (95:5) | -- | -- |
| 26-28 | Pet. Ether: Chloroform (93:7) | -- | -- |
| 29-35 | Pet. Ether: Chloroform (92:8) | -- | -- |
| 36-42 | Pet. Ether: Chloroform (90:10) | -- | -- |
| 43-54 | Pet. Ether: Chloroform (90:10) | Single spot | A (light yellow) |
| 55-64 | Pet. Ether: Chloroform (90:1) | -- | -- |
| 65-73 | Pet. Ether: Chloroform (87:13) | -- | -- |
| 74-81 | Pet. Ether: Chloroform (85:15) | -- | -- |
| 82-83 | Pet. Ether: Chloroform (83:17) | -- | -- |
| 84-86 | Pet. Ether: Chloroform (80:20) | -- | -- |
| 87-91 | Pet. Ether: Chloroform (70:30) | -- | -- |
| 92-113 | Pet. Ether: Chloroform (50:50) | -- | -- |
| 114-127 | Pet. Ether: Chloroform (40:60) | -- | -- |
| 128 – 133 (distilled) | Pet. Ether: Chloroform (40:60) | Single spot | B (faint green) |
| 134-151 | Pet. Ether: Chloroform (40:60) | -- | -- |
| 152 – 169 (distilled) | Pet. Ether: Chloroform (30:70) | -- | -- |
| 170 – 182 (distilled) | Pet. Ether: Chloroform (20:80) | -- | -- |
| 183-199(distilled) | Pet. Ether: Chloroform (20:80) | Single spot | C (yellow) |
| 200 – 210 (distilled) | Pet. Ether: Chloroform (10:90) | -- | -- |
| 211 – 232 (distilled) | Pet. Ether: Chloroform (5:95) | -- | -- |
| 233 – 247 (distilled) | Pet. Ether: Chloroform (97:3) | -- | -- |
| 248 – 253 (distilled) | Pet. Ether: Chloroform (99:1) | -- | -- |
| 254 – 256 (distilled) | Chloroform (100%) | -- | -- |
| 257 – 262 (distilled) | Chloroform: methanol (90:10) | Single spot | D (orange) |
| 263 – 265 (distilled) | Chloroform: methanol (85:15) | -- | -- |
| 266 – 268 (distilled) | Chloroform: methanol (85:15) | Single spot | E (light orange) |
| 269 – 271 (distilled) | Chloroform: methanol (85:15) | -- | -- |
| 272 – 281 (distilled) | Chloroform: methanol (85:15) | Single spot | F (yellow) |
| 282 – 284 (distilled) | Chloroform: methanol (70:30) | -- | -- |
| 285 – 287 (distilled) | Chloroform: methanol (70:30) | -- | -- |
| 288 – 292 (distilled) | Chloroform: methanol (60:40) | -- | -- |
| 293 – 297 (distilled) | Chloroform: methanol (50:50) | -- | -- |
| 298 – 302 (distilled) | Chloroform: methanol (30:70) | -- | -- |
| 303 – 307 (distilled) | Chloroform: methanol (20:80) | -- | -- |
| 308 – 320 (distilled) | Methanol (100%) | -- | -- |

Qualitative Analysis of various Fractions**1) Fraction A****a. IR Spectrum**

| Sr. No. | Peaks | Functional Structural form | Group Names |
|---------|---------|----------------------------|---------------------------------|
| 1. | 2921.23 | C-H | Alkanes and aliphatic molecules |
| 2. | 2851.78 | C-H | Alkanes and aliphatic molecules |
| 3. | 1711.88 | C=O | Ketone |
| 4. | 1462.76 | C=C | Aromatic ring |

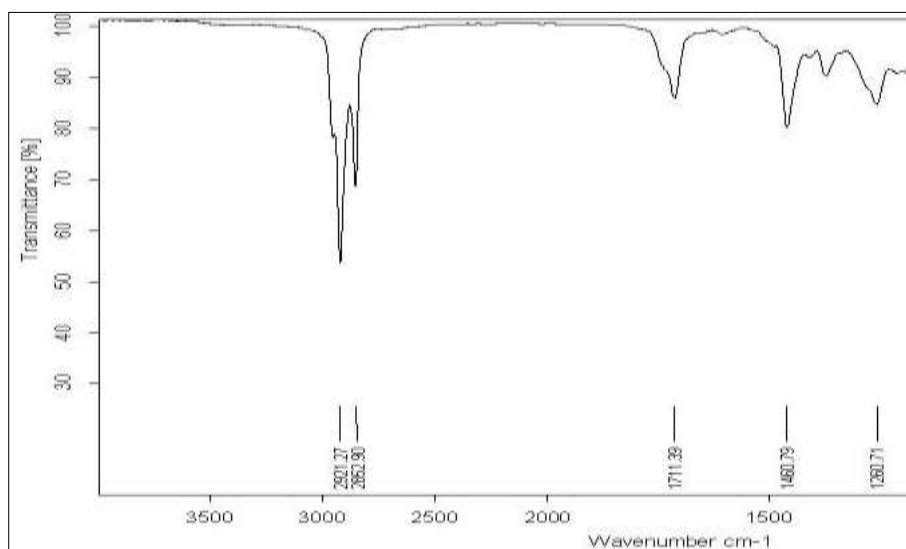


2) **Fraction B**
a. **IR Spectrum**



| Sr. No. | Peaks | Functional Structural form | Group Names |
|---------|---------|----------------------------|--------------------------------|
| 1. | 2921.79 | C-H | Alkanes and aromatic molecules |
| 2. | 2853.12 | C-H | Alkanes and aromatic molecules |
| 3. | 1732.43 | C=O | Ketones |
| 4. | 1461.71 | C=C | Aromatic ring |
| 5. | 1260.79 | C-O | Alcoholic |

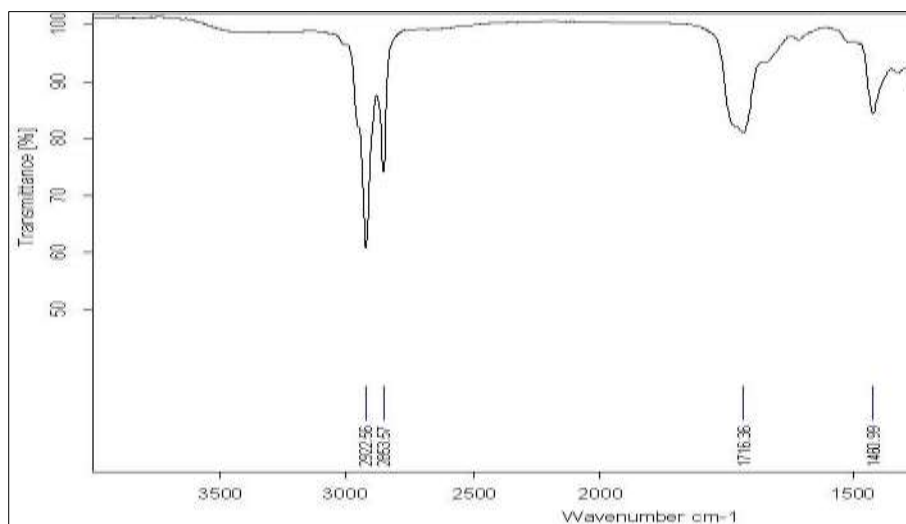
3) **Fraction C**
a. **IR Spectrum**



| Sr. No. | Peaks | Functional Structural form | Group Names |
|---------|---------|----------------------------|--------------------------------|
| 1. | 2921.27 | C-H | Alkanes and aromatic molecules |
| 2. | 2852.90 | C-H | Alkanes and aromatic molecules |
| 3. | 1711.39 | C=O | Ketone |
| 4. | 1460.79 | C=C | Aromatic ring |

4) Fraction D

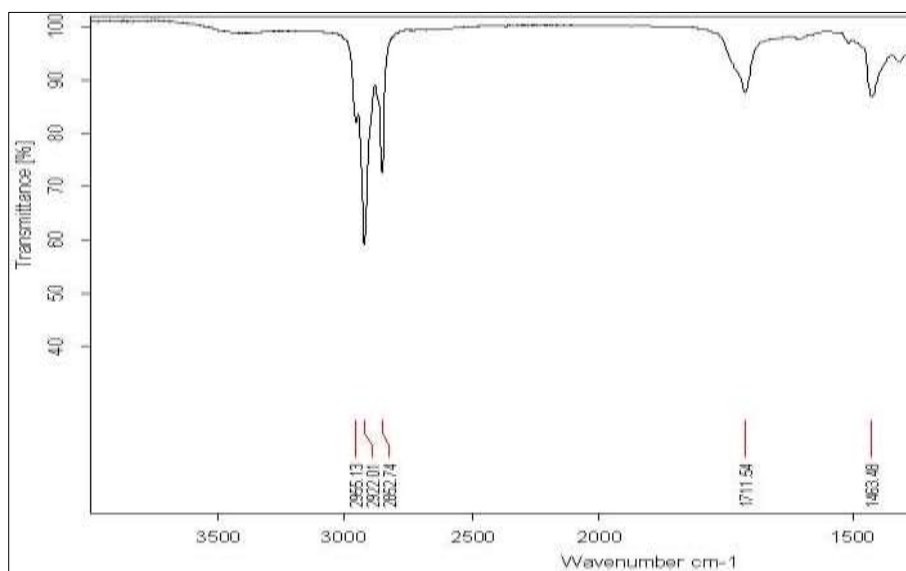
a. IR Spectrum



| Sr. No. | Peaks | Functional Structural form | Group Names |
|---------|---------|----------------------------|--------------------------------|
| 1. | 2922.56 | C-H | Alkanes and aromatic molecules |
| 2. | 2853.57 | C-H | Alkanes and aromatic molecules |
| 3. | 1716.36 | C=O | Ketone |
| 4. | 1460.99 | C=C | Aromatic ring |

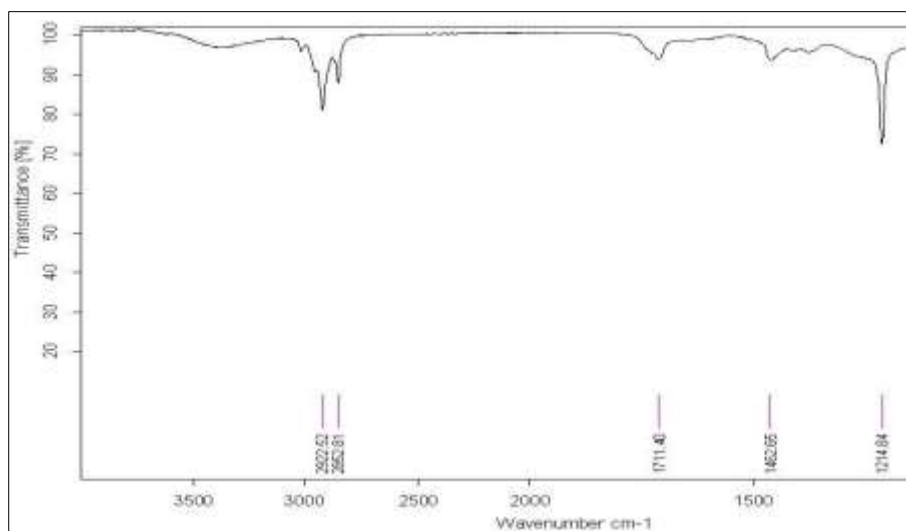
Fraction E

a. IR Spectrum



| Sr. No. | Peaks at | Functional | Group |
|---------|----------|-----------------|--------------------------------|
| | | Structural form | Names |
| 1. | 2955.13 | C-H | Alkanes and aromatic molecules |
| 2. | 2922.01 | C-H | Alkanes and aromatic molecules |
| 3. | 2852.74 | C-H | Alkanes and aromatic molecules |
| 4. | 1711.54 | C=O | Ketone |
| 5. | 1463.48 | C-H | Alkanes |

Fraction F a. IR Spectrum



| Sr. No. | Peaks | Functional | Group |
|---------|---------|-----------------|--------------------------------|
| | | Structural form | Names |
| 1. | 2922.52 | C-H | Alkanes and aromatic molecules |
| 2. | 2852.81 | C-H | Alkanes and aromatic molecules |
| 3. | 1711.40 | C=O | Ketone |
| 4. | 1462.65 | C=C | Aromatic ring |
| 5. | 1214.84 | C-O | Alcoholic |