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Wolbachia: An evolutionary way to combat mosquito borne disease and the challenges in success of the strategy

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Abstract

Wolbachia is an intracellular bacterium that is found in many insects and very commonly in Dipterans. It is now being extensively studied as it aids in suppression of mosquito populations. Mosquitoes are causative agents of many life-threatening diseases (dengue, malaria, chikungunya fever, Zika virus fever, yellow fever, West Nile fever, Japanese encephalitis and put immense burden on health systems of many countries. *Wolbachia* is known to block replication of several arboviruses. However, the exact mechanism how it does so is still under progress. It is transmitted vertically & alters host biology in several ways, one of the keys is cytoplasmic incompatibility, a trait which is used to suppress mosquito population by artificially introducing them into host. Here, in this review we will try to unravel the story of *Wolbachia* by looking into its role in manipulating host immune system and also how it became a miraculous weapon for controlling mosquito borne diseases, and also the challenges related to implication of this system.

Keywords: Wolbachia, Aedes, cytoplasmic incompatibility, dengue, chikungunya, malaria, vector

Introduction

Suffering and mortality caused by mosquitoes is quite a common thing which in itself doesn't need much introduction. Malaria is a parasitic infection transmitted by Anopheline mosquitoes. It causes an estimated 219 million cases globally, and results in more than 400,000 deaths every year. Most of the deaths found in children were under the age of 5 years ^[1]. Dengue is the most prevalent and deadliest viral infection transmitted by *Aedes* mosquitoes ^[2]. More than 3.9 billion people in over 129 countries are at risk of contracting dengue, with an estimated 96 million symptomatic cases and an estimated 40,000 deaths every year ^[2]. Other viral diseases transmitted by these vectors include chikungunya fever, Zika virus fever, yellow fever, West Nile fever, Japanese encephalitis ^[1-3]. Virus is not the only pathogen transmitted by this genus of mosquito lymphatic filariasis is the other example of disease spread by *Ae. polynesiensis* in South Pacific specific features of this virus may have been the failure of drugbased control programme in that region ^[4, 5].

Although a vaccine has been developed for yellow fever but Dengue lacks an effective vaccine. Dengue control largely focuses on control of vector spreading this disease as there are no therapeutic or prophylactic drug against and effective vaccine. Current mosquito control programmes are struggling to address elimination of complete breeding sites of container breeder *Aedes*. There are comprehensive vector management is still long way to go in achieving the targets ^[6-9]. Although mass awareness could help in such issues but even if one household forgets to follow the measures breeding of mosquito continues hence, failing the purpose. Adulticides have also very limited effectiveness, which increases the risk of resistance development and also bed nets do not provide a solution to day-biting mosquitoes. In the changed urban settings to cope up vector control in high population density areas with environmental safety the present system of vector control may not be adequate there is a need to revise and incorporate new tools that are also suitable for environment. High population density, uncontrolled travels, inadequate manpower for surveillance, lack of integrated approach etc.

These seeks a new approach to combat the disease spread by mosquitoes and were the main reason for development of new genetic based approaches. Genetic based approaches solve many problems like they are extremely species – specific i.e., other populations of insects or mosquitoes will not be affected hence environmentally safe. However, drawback of genetic based approach is that it is not helpful for the diseases in which pathogen is spread by multiple species. One such genetic approaches uses *Wolbachia* an intracellular bacterium which once infect mosquito has property of causing cytoplasmic incompatibility and hence helping in the population control of that particular species of mosquito ^[4, 10, 11].

Mechanism of Wolbachia action

The bacterial endosymbiont *Wolbachia pipientis* more commonly referred to as *Wolbachia* was previously used to protect flies from virus induced mortality ^[12]. Later three independent studies presented its scope to inhibit pathogens like, *Plasmodium*, Chikungunya, Dengue and filarial nematode in *Ae. aegypti* ^[13-15]. Further, it was shown to inhibit Zika, West Nile, bluetongue virus and yellow fever ^[16-17]. Pathogen interference by *Wolbachia* can be classified into three types as tabulated in table 1.

Table 1:	Three	types of	pathogen	interference	mechanisms

Pathogen Interference						
In the trans	In mosquito or flies with	Use of transient				
infected mosquito	native Wolbachia	somatic Wolbachia to				
[10, 18, 19]	infection [20-23]	inhibit pathogen.[24-26]				

But, the most intriguing phenotype that is induced by Wolbachia is cytoplasmic incompatibility which results when a Wolbachia infected male mates with female that either is uninfected or infected with some other strain of Wolbachia leads to sterile mating leading to embryonic death [27]. Although the mechanism behind cytoplasmic incompatibility is unknown one of the accepted hypotheses is that when male infected with Wolbachia mates with either uninfected female or female infected with some other strain, it leads to condensation of paternal chromosome, which is then lost in early embryonic development. However, when the mating takes place between partners infected with same Wolbachia strain the paternal material somehow restores functionality resulting in successful embryonic development ^[28]. When Wolbachia is infected in female mosquito it is vertically inherited i.e., will always be passed on from one generation to the next but if only males have Wolbachia then there is sterile mating with females i.e., no successful embryonic development

So, this mechanism in turn gives two strategies to use against mosquito borne disease

- 1. **Population suppression:** If we release males infected with *Wolbachia* in environment, sterile mating's will ultimately lead to suppression of mosquito population ^[29, 30].
- 2. **Population replacement:** Release of female mosquitoes infected with *Wolbachia* along with infected males, thus only chances of successful mating will be that between uninfected mosquitoes or between infected mosquitoes which over the period of generation will keep on increasing and thus resulting in invasion of *Wolbachia* into population ^[31-34-,10].

Challenges faced in this system

One of the challenges among other is that *Wolbachia* is naturally not present in mosquitoes hence mosquito needs to be transfected in lab only.

Does Wolbachia affect life history trait of mosquito?

When infecting mosquito or any other organism with organism that is not native to it the prime question is whether or not will this affect the life-history of organism concerned. Fortunately, in the case of infecting mosquito with *Wolbachia* there was no impact on inter and intra-specific competition. Studies on development time showed controversial result hence this aspect can be said to be dependent on mosquito species and *Wolbachia* strain being used ^[35]. However, presence of *Wolbachia* reduces the starvation tolerance in *Ae. aegypti* ^[36].

Can *Wolbachia* infected mosquito population successfully invade the wild population?

Population density of wild mosquitoes pose a threat to the invasion of *Wolbachia* infected mosquitoes ^[37, 38]. Not only the same species that pose threat to *Wolbachia* infected mosquito invasion but also other species of mosquitoes that share the breeding ground also forms an obstacle in invasion. So, before releasing the mosquitoes infected with *Wolbachia* in environment the estimation of population size of different mosquitoes that share the same breeding ground needs to be done. So, that the optimum size of population that needs to be released in that particular site can be ascertained ^[38-43]. Researchers have all but obliterated populations of the world's most invasive mosquito species - the Asian tiger mosquito (*Aedes albopictus*) - on two islands in the Chinese city of Guangzhou ^[44].

Is it good to release a self-sustaining genetic system into environment?

For any self-sustaining genetic system it is a must that evolutionary responses do not compromise its effectiveness i.e., an initial ability of system to curb the thing for which it is made in this case fertility of mosquito over generation then only it can be called as evolutionary successful genetic model ^[45] While the strategy of *Wolbachia* provides a promising result its long-term effects still need to be studied, like for how many generations it will affect or whether this approach can lead to development of resistant strains of virus ^[46-49]. According to a study, wMelPop infected.

Ae. aegypti require human blood to lay viable eggs in contrast to normal uninfected one. This study suggests that introducing *Wolbachia* might give selection to human biting preference in mosquito ^[50].

These issues are major concerns to look at before releasing this self- sustaining system in environment, as this decision can surely have repercussions. On the one hand *Wolbachia* vouch for being a natural remedy for this situation on the other hand it can also throwback surprises on evolutionary front.

Conclusion

Wolbachia has come up with having lots of potential to be an alternate approach in suppressing mosquito-borne diseases. Its virus blocking property has made people invest more in research relating to *Wolbachia*.

However, before applying this on field lot of research still

needs to be done as no one would ever want a laboratory grown thing to backfire. Despite of the fact that we still don't have all the knowledge how *Wolbachia* works to block the transmission of virus it is giving us positive results. So, in order to utilize *Wolbachia* to its full potential mechanism behind it needs to be understood more clearly.

References

- https://www.who.int/news-room/fact-sheets/detail/vectorborne-diseases
- 2. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, *et al*. The global distribution and burden of dengue. Nature. 2013;496(7446):504-7.
- 3. WHO. Global strategy for dengue prevention and control 2012-2020. WHO, 2012.
- 4. Chambers EW, Hapairi L, Peel BA, Bossin H, Dobson SL. Male mating competitiveness of a *Wolbachia*introgressed *Aedes polynesiensis* strain under semi-field conditions. PLoSNegl Trop Dis. 2011;5(8):e1271.
- O'Connor L, Plichart C, Sang AC, Brelsfoard CL, Bossin HC, Dobson SL. Open release of male mosquitoes infected with a *Wolbachia* biopesticide: field performance and infection containment. PLoSNegl Trop Dis. 2012;6:e1797.
- Nalini Mishra, Nidhi Krishna Shrivastava, Ajay Nayak, Himmat Singh. *Wolbachia*: A prospective solution to mosquito borne diseases. International Journal of Mosquito Research. 2018;5(2):01-08.
- 7. Egger JR, Ooi EE, Kelly DW, Woolhouse ME, Davies CR, Coleman PG. Reconstructing historical changes in the force of infection of dengue fever in Singapore: implications for surveillance and control. Bull World Health Organ. 2008;86:187-96.
- Ooi EE, Goh KT, Gubler DJ. Dengue prevention and 35 years of vector control in Singapore. Emerg Infect Dis. 2006;12:887-93.
- 9. National Environment Agency. Campaign against dengue, 2012. http://www.dengue.gov.sg/
- 10. Xi Z, Khoo CC, Dobson SL. *Wolbachia* establishment and invasion in an *Aedes aegypti* laboratory population. Science. 2005;310:326-8.
- 11. Xi Z, Khoo CC, Dobson SL. Interspecific transfer of *Wolbachia* into the mosquito disease vector *Aedes albopictus*. Proc Biol Sci. 2006;273:1317-22.
- 12. Hedges LM, Brownlie JC, O'Neill SL, Johnson KN, *Wolbachia* and virus protection in insects. Science, 2008;322:702.
- 13. Bian G, Xu Y, Lu P, Xie Y, Xi Z. The endosymbiotic bacterium *Wolbachia* induces resistance to dengue virus in *Aedes aegypti*. PLoSPathog. 2010;6:e1000833.
- 14. Kambris Z, Cook PE, Phuc HK, Sinkins SP. Immune activation by life-shortening *Wolbachia* and reduced filarial competence in mosquitoes. Science. 2009;326:134-136.
- 15. Moreira LA, Iturbe-Ormaetxe I, Jeffery JA, Lu G, Pyke AT, Hedges LM, *et al.* A *Wolbachia* symbiont in *Aedes aegypti* limits infection with dengue, Chikungunya, and *Plasmodium.* Cell. 2009;139:1268-1278.
- Shaw AE, Veronesi E, Maurin G, Ftaich N, Guiguen F, Rixon F, *et al. Drosophila melanogaster* as a model organism for bluetongue virus replication and tropism. J Virol. 2012;86:9015-9024.
- 17. Dutra HL, Rocha MN, Dias FB, Mansur SB, Caragata

EP, Moreira LA. *Wolbachia* blocks currently circulating zika virus isolates in Brazilian *Aedes aegypti* mosquitoes. Cell Host Microbe. 2016;19:771-774.

- McMeniman CJ, Lane RV, Cass BN, Fong AW, Sidhu M, Wang YF, *et al.* Stable introduction of a lifeshortening *Wolbachia* infection into the mosquito *Aedes aegypti*. Science. 2009;323:141-144.
- 19. Walker T, Johnson PH, Moreira LA, Iturbe-Ormaetxe I, Frentiu FD, McMeniman CJ, *et al*. The wMelWolbachia strain blocks dengue and invades caged *Aedes aegypti* populations. Nature. 2011;476:450-453.
- 20. Baton LA, Pacidonio EC, Goncalves DS, Moreira LA. wFlu: characterization and evaluation of a native *Wolbachia* from the mosquito *Aedes fluviatilis* a potential vector control agent. PLoS One. 2013;8:e59619.
- Tsai KH, Huang CG, Wu WJ, Chuang CK, Lin CC, Chen WJ. Parallel infection of Japanese encephalitis virus and *Wolbachia* within cells of mosquito salivary glands. J Med. Entomol. 2006;43:752-756.
- 22. Panteleev D, Goriacheva II, Andrianov BV, Reznik NL, Lazebnyi OE, Kulikov AM. The endosymbiotic bacterium *Wolbachia* enhances the nonspecific resistance to insect pathogens and alters behavior of *Drosophila melanogaster*. Genetika. 2007;43:1277-1280.
- 23. Micieli MV, Glaser RL. Somatic *Wolbachia* (Rickettsiales: Rickettsiaceae) levels in *Culex quinquefasciatus* and *Culex pipiens* (Diptera: Culicidae) and resistance to West Nile virus infection. J Med. Entomol. 2014;51:189-199.
- 24. Hughes GL, Koga R, Xue P, Fukatsu T, Rasgon JL. *Wolbachia* infections are virulent and inhibit the human malaria parasite *Plasmodium falciparum* in anopheles gambiae. PLoS Pathog. 2011;7:e1002043.
- Hughes GL, Vega-Rodriguez J, Xue P, Rasgon JL. Wolbachia strain wAlbB enhances infection by the rodent malaria parasite *Plasmodium berghei*in *Anopheles* gambiae mosquitoes. Appl. Environ. Microbiol. 2012;78:1491-1495.
- Kambris Z, Blagborough AM, Pinto SB, Blagrove MS, Godfray HC, Sinden RE, *et al. Wolbachia* stimulates immune gene expression and inhibits plasmodium development in *Anopheles gambiae*. PLoS Pathog, 2010, 6.
- Hoffmann AA, Turelli M. Cytoplasmic incompatibility in insects. In: O'Neill SL, Hoffmann AA, Werren JH. (Eds.), Influential Passengers: Inherited Microorganisms and Arthropod Reproduction. Oxford University Press, Oxford, 1997, pp42-80.
- Poinsot D, Charlat S, Mercot H. On the mechanism of Wolbachia-induced cytoplasmic incompatibility: confronting the models with the facts. Bioessays. 2003;25:259-265.
- 29. Laven H. Eradication of *Culex pipiens fatigans* through cytoplasmic incompatibility. Nature. 1967;216:383-384.
- O'Connor L, Plichart C, Sang AC, Brelsfoard CL, Bossin HC, Dobson SL. Open release of male mosquitoes infected with a *Wolbachia* biopesticide: field performance and infection containment. PLoSNegl. Trop. Dis. 2012;6:e1797.
- Bian G, Joshi D, Dong Y, Lu P, Zhou G, Pan X, et al. Wolbachia invades Anopheles stephensi populations and induces refractoriness to Plasmodium infection. Science. 2013a;340:748-751.

- Curtis CF, Sinkins SP. *Wolbachia* as a possible means of driving genes into populations. Parasitology. 1998;116(Suppl.):S111-S115.
- Hoffmann AA, Montgomery BL, Popovici J, Iturbe-Ormaetxe I, Johnson PH, Muzzi F, *et al.* Successful establishment of *Wolbachia* in *Aedes* populations to suppress dengue transmission. Nature. 2011;476:454-457.
- Turelli M, Hoffmann AA. Microbe-induced cytoplasmic incompatibility as a mechanism for introducing transgenes into arthropod populations. Insect Mol. Biol. 1999;8:243-255.
- 35. De Oliveira S, Villela DAM, Dias FBS, Moreira LA, Maciel de Freitas R. How does competition among wild type mosquitoes influence the performance of *Aedes aegypti* and dissemination of *Wolbachia pipientis*? PLoSNegl. Trop. Dis. 2017;11:e0005947. DOI: 10.1371/journal.pntd.0005947.
- Ross PA, Endersby NM, Hoffmann AA. Costs of three Wolbachia infections on the survival of Aedes aegypti larvae under starvation conditions. PLoSNegl Trop Dis. 2016;10:e0004320.
- 37. Ritchie SA, Montgomery BL, Hoffmann AA. Novel estimates of *Aedes aegypti* (Diptera: Culicidae) population size and adult survival based on *Wolbachia* releases. J Med Entomol. 2013;50:624-631.
- Garcia GA, Santos LM, Villela DA, Maciel-de-Freitas R. Using Wolbachia releases to estimate Aedes aegypti (Diptera: Culicidae) population size and survival. PLoS One. 2016;11:e0160196.
- Hancock PA, White VL, Ritchie SA, Hoffmann AA, Godfray HC. Predicting *Wolbachia* invasion dynamics in *Aedes aegypti* populations using models of densitydependent demographic traits. BMC Biol. 2016;14(1):96.
- Irving-Bell RJ, Okoli EI, Diyelong DY, Lyimo EO, Onyia OC. Septic tank mosquitoes: competition between species in central Nigeria. Med Vet Entomol. 1987;1(3):243-50.
- 41. Lourenço-de-Oliveira R, Vazeille M, De Filippis AM, Failloux AB. *Aedes aegypti* in Brazil: genetically differentiated populations with high susceptibility to dengue and yellow fever viruses. Trans R Soc Trop Med Hyg. 2004;98(1):43-54.
- 42. Villela DAM, Garcia GA, Maciel-de-Freitas R. Novel inference models for estimation of abundance, survivorship and recruitment in mosquito populations using mark-release-recapture data. PLoSNegl Trop Dis. 2017;11:e0005682.
- 43. Dutra HL, dos Santos LMB, Caragata EP, Silva JB1, Villela DA, Maciel-de-Freitas R, *et al.* From lab to field: the influence of urban landscapes on the invasive potential of Wolbachia in Brazilian *Aedes aegypti* mosquitoes. PLoSNegl Trop Dis. 2015;9:e0003689.
- 44. Lu P, Bian G, Pan X, Xi Z. *Wolbachia* induces densitydependent inhibition to dengue virus in mosquito cells. PLoSNegl Trop Dis. 2012;6:e1754.
- 45. Zheng X. *et al.* Nature, 2019. https://doi.org/10.1038/s41586-019-1407-9.
- 46. Scott TW, Chow E, Strickman D, Kittayapong P, Wirtz RA, Lorenz LH, *et al.* Blood-feeding patterns of *Aedes aegypti* (Diptera: Culicidae) collected in a rural Thai village. J Med Entomol. 1993;30:922-7.
- 47. Siriyasatien P, Pengsakul T, Kittichai V, Phumee A,

Kaewsaitiam S, Thavara U, *et al.* Identification of blood meal of field caught *Aedes aegypti* (L.) by multiplex PCR. Southeast Asian J Trop Med Public Health. 2010;41:43-7.

- 48. Valerio L, Marini F, Bongiorno G, Facchinelli L, Pombi M, Caputo B, *et al.* Host-feeding patterns of *Aedes albopictus* (Diptera: Culicidae) in urban and rural contexts within Rome province, Italy. Vector Borne and Zoonotic Dis. 2010;10:291-4.
- 49. Barrera R, Bingham AM, Hassan HK, Amador M, Mackay AJ, Unnasch TR. Vertebrate hosts of *Aedes aegypti* and *Aedes mediovittatus* (Diptera: Culicidae) in rural Puerto Rico. J Med Entomol. 2012;49:917-21.
- 50. McMeniman CJ, Hughes GL, O'Neill SL. A *Wolbachia* symbiont in *Aedes aegypti* disrupts mosquito egg development to a greater extent when mosquitoes feed on nonhuman versus human blood. J Med Entomol. 2011;48:76-84.