



ISSN: 2348-5906
CODEN: IJMRK2
IJMR 2018; 5(6): 10-14
© 2018 IJMR
Received: 03-09-2018
Accepted: 04-10-2018

Bhubaneshwari Devi Moirangthem
Laboratory of Entomology,
P.G. Department of Zoology
D.M. College of Science, Imphal,
Manipur, India

Salam Noren Singh
Department of Statistics, Imphal
College Imphal, Manipur, India

Dhananjay Chingambam Singh
Laboratory of Entomology,
P.G. Department of Zoology
D.M. College of Science, Imphal,
Manipur, India

Comparative studies of three potent bioagent against mosquito larvae

Bhubaneshwari Devi Moirangthem, Salam Noren Singh and Dhananjay Chingambam Singh

Abstract

Mosquitoes are menace to human for centuries with their capability to spread dreaded diseases like malaria, dengue, filariasis etc. the control or managing them is the first priority of the humans ever since the cause of such diseases has been revealed. The usual ways to control include the spraying of insecticides or fumigation. Utilization of predacious insect larvae as well as the mosquito larvae are eco-friendly, sustainable and cost effective methods for the purposes. In the present study three insect larvae viz. Odonata (*Sympatrum* sp.), Hemiptera (*Diplonychus* sp.) and Diptera (*Lutzia tigris*) were reared in the laboratory to check their efficacy on controlling the mosquito larvae efficiently. The larvae feeding them were the *Culex* species larvae. To find out the insect consuming larvae significantly more than others Turkey's HSD for 3 means and 2 means are calculated and is found to be 61.19 and 48.79 respectively. Comparing the differences between the means, all means are found to be significantly different. The insect consuming larvae significantly than others is Odonata. In the same way, to find out the instar of the larvae at which insects consume them in significant amount Turkey's HSD is calculated for 4, 3 and 2 means and they are 79.77, 70.66 and 56.34 respectively. Comparing the differences in the means with these values, it is found that insects consume larvae at 4th instar significantly more than the larvae at other stages. Out of the three larvae for managing or even controlling the mosquito larvae, *Lutzia* larvae might be the most efficient candidate for releasing in wild to control for they can survive and well adapted to the any habitats whether polluted or non-polluted but the Odonata and Hemiptera prefer somewhat non polluted aquatic habitats having moderate amount of dissolved oxygen. The main focus of the study is to select a bioagent that has no habitat boundary or constrain and nature of aquatic habitat. Hence the efficient candidate for controlling the mosquitoes of a particular habitat will be to utilize nature's best weapon against the mosquitoes that is *Lutzia tigris*.

Keywords: Predacious insects, Odonata, Hemiptera, *Lutzia tigris*, mosquito larvicides, management

1. Introduction

Mosquitoes are menace to human for centuries with their capability to spread dreaded diseases like malaria, dengue, filariasis etc. The control or managing them is the first priority of the humans ever since the cause of such diseases has been revealed. Efforts of managing in worldwide yet jet-linked world remains on the edge of resurgence and out- breaks of old and new mosquito-borne disease epidemics^[1, 2]. Biological control is most suitable in this context as few attempt for introduction of biological agents like larvivorous fishes i.e. gambia and guppy have been successful^[3], bacteria^[4, 5], fungi^[6, 7] and predatory mosquitoes of subgenera *Mucidus*^[8], *Culex fuscans*, *Toxorhynchites* spp.^[9, 10] to control *Aedes aegypti* and *Culex quinquefasciatus* larvae^[11]. Utilizing biological organisms to control mosquito larvae is not only eco-friendly, but constitutes a means by which more effective and sustainable control can be achieved.

Among the predacious insects promising as biological control are Hemipteran, Odonata and Diptera (*Lutzia tigris*) predators which are cosmopolitans and locally available. The backswimmers (Family: Notonectidae) are the most common bugs preying upon mosquito larvae, important factor in reducing immature mosquito population and considered promising in mosquito control. The role of hemipteran predators in controlling mosquito larvae has been recognized since 1939 in New Zealand, when stock troughs with *Anisops assimilis* were found to be free of mosquitoes whereas puddles in depressions surrounding the troughs contained mosquitoes^[12].

Correspondence
Salam Noren Singh
Department of Statistics, Imphal
College Imphal, Manipur, India

Bay [13] found that almost 100% of mosquito emergence was prevented in field-situated, screened, 100 gallon fiberglass tubs with one square meter of water surface and *Notonecta unifasciata* compared to more than 12000 adult mosquitoes emerged from the control tubs. For instance, emergent vegetation in ponds and other water bodies provide partial protection for mosquito immatures. This effect was experimentally investigated and confirmed by Shaalan [14] and Shaalan *et al.* [15] whereas predation potential of *Anisops* and *Diplonychus* bugs was significantly reduced by the presence of vegetation. Although the costs of colonization and mass production, coupled with the logistics of distribution, handling and timing of release at the appropriate breeding site, impede the use of notonectids in mosquito control [16], results of a recent study for mass rearing and egg release of the predatory backswimmer *Buenoa scimitar* for the biological control of *Cx. quinquefasciatus* were impressive [17]. The dragonfly larvae of *Trithemis annulata scortecii* were intense and active predators when used to control mosquito larvae, especially *Anopheles pharoensis*, in irrigation channels in Gezira Province, Sudan [18]. Bay [13] reported that dragonfly larvae are known to prey heavily on bottom feeder mosquitoes like *Aedes* larvae. Sebastian *et al.* [19] found that complete elimination of all *Ae. aegypti* larvae and pupae were achieved between day 4 and 9 depending on the density of aquatic stages of mosquitoes present per container when dragonfly larva, *Labellula sp.*, was used, Sebastian *et al.* [20] conducted a pilot field study, involving periodic augmentative release of predaceous larvae of a dragonfly, *Crocothemis servilia*, to suppress *Ae. aegypti* during the rainy season in Yangon, Myanmar. Chatterjee *et al.* [21] found that significant decrease in *An. subpictus* larval density in dipper samples was observed 15 days after the introduction of *Brachytron pretense* dragonfly larvae in concrete tanks under field conditions in India. Similarly, the larvae of 5 odonate species *Aeshna flavifrons*, *Coenagrion kashmirum*, *Ischnura forcipata*, *Rhinocypha ignipennis* and *Sympetrum durum* in semifield conditions in West Bengal, India, significantly lowered the mosquito larval density in dipper samples after 15 days from the introduction, followed by a significant increase of larval mosquito density after 15 days from the withdrawal of the larva [22]. These results [18, 19, 20, 21, 22] are suggestive of

the use of odonate larvae as potential biological agent in regulating the larval population of mosquito vectors. Breene *et al.* [23] found no mosquito larvae in the gut of the larvae of the damselfly *Enallagma civile*. Biology, colonization and potential of *Toxorhynchites* mosquitoes as a biological control agent of vector mosquitoes are fully covered by Collins and Blackwell [24] while Garcia [25] discussed the difficulties associated with such methodologies which prevent more widespread utilization of arthropod predators. In addition to *Toxorhynchites* mosquitoes, the predaceous characters of *Culex* (Subgenus *Lutzia*) mosquitoes were reviewed by Pal and Ramalingam [26]. Lacey and Orr [27] limited their discussion to insect predators that are used as biological control agents in integrated vector control to *Notonecta* and *Toxorhynchites* species. Mogi [28] reviewed insects and invertebrate predators based on adult, egg, larval and pupal mosquito predation beside possibilities of using such predators for mosquito control. Quiroz-Martinez *et al.* [29] discussed the arthropods (insects, mites and spiders) that prey on mosquito larvae and considerations for the success of these predators in mosquitoes' biological control programs.

The present article reports the predation on mosquito, particularly larvae using three predacious insects viz. Odonata, Hemiptera and larval predacious mosquito *Lutzia tigris* in the laboratory by feeding on *Culex* larvae. The effectiveness of three are quite similar but regarding the field released and adaptability to different habitat the

2. Materials and Methods

Two predacious larval forms of insects and one mosquito larva known for its predacious habits were selected for the study. They were identified in accordance with the available references and expertise. For feeding *Culex* larvae were collected from surrounding habitat. For easy observation and feeding small washing dishes were used for the experiments. The mosquito larvae were counted before putting into and duration and stage of the insect were thoroughly recorded throughout the experiment.

To find out the insect consuming larvae significantly more than others Turkey's HSD for 3 means and 2 means were calculated.

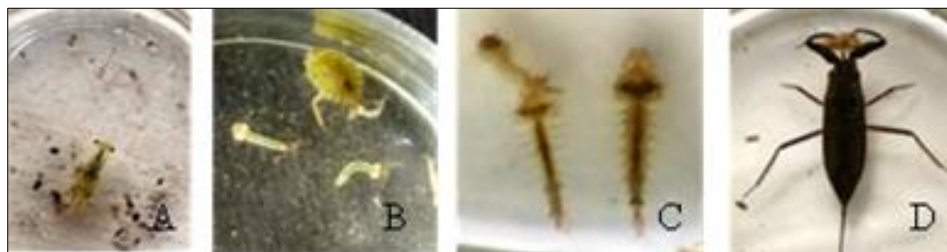


Fig 1: The predacious immature insects used in the present study (Odonata-A, Hemiptera –B and *Lutzia tigris* mosquito predating on the *Culex* larva-C). *Laccotrephes ruber* predate on the Hemipteran and Odonata nymphs (D) neutralizing the ill effect of the pathogens.

3. Results

The predacious insects and larvae, in the present study effectively consumed or killed the mosquito larvae. Above all these insects were lastly killed by *Laccotrephes ruber* predate on the Hemipteran and Odonata nymphs neutralizing the ill effect of the pathogens (fig. 1). The Odonata and *Lt. tigris* consumed whole of the mosquito larvae while the hemipteran larvae killed the mosquito larvae by sucking

fluids. The efficacy of the predators is shown in figure 2. The efficacy is tested statistically comparing the three insects.

3.1 Statistical results

To find out the insect consuming larvae significantly more than others Turkey's HSD for 3 means and 2 means were calculated and was found to be 61.19 and 48.79. Comparing the differences between the means, all means were found to

be significantly difficult. The insect consuming larvae significantly than others was Odonata. In the same way, to find out the instar of the larvae at which insects consume them in significant amount Turkey's HSD was calculated for 4, 3

and 2 means and they were 79.77, 70.66 and 56.34 respectively. Comparing the differences in the means with these values, it was found that insects consume larvae at 4th instar significantly more than the larvae at other stages.

Table 1: 24-hour observation on consumption pattern of three different insects viz., Odonata, Hemiptera and Dipteran on different stages (instars) of mosquito larvae.

Instar of Insects	Odonata				Hemiptera				Diptera			
	Replication				Replication				Replication			
	1	2	3	4	1	2	3	4	1	2	3	4
1 st												
2 nd												
3 rd												
4 th												

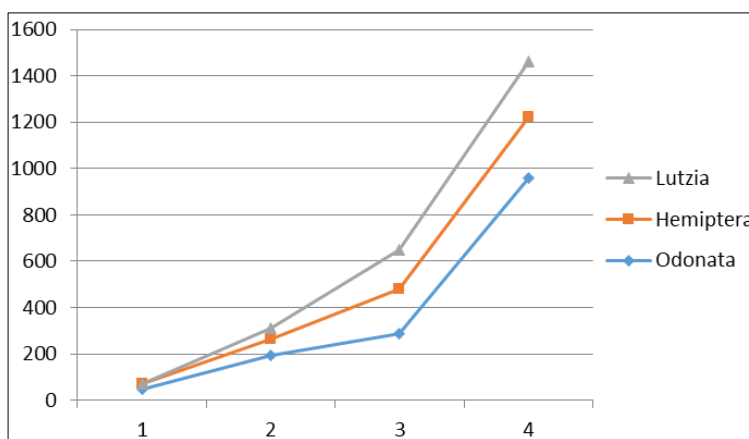


Fig 2: The comparative efficiencies of three predacious insects and data were extracted from Table 1 (Data not shown).

Analysis of variance two-way classification with 4 observations per cell:

The null hypotheses to be tested are:

- H01: the different types of insects consume larvae equally.
- H02: there is no significant difference in the consumption of larvae in respect of different instars
- H03: there are no significant influences of the different types of insects on different instars of larvae.

For testing the above three hypotheses we use analysis of variance two-way classification with four observations per cell and its ANOVA was given as below:

ANOVA of two-way classification

$$\text{Interaction SS} = \text{Between Groups SS} - (\text{SS Instar} + \text{SS Insects})$$

$$= 1972506.67 - (1164912.00 + 426834.67)$$

$$= 1972506.67 - 1591746.67$$

$$= 380760.00$$

or

$$\text{Interaction SS} = \{(360^2/4 + 288^2/4 + 0^2/4 + \dots + 1896^2/4)\} - \text{CF} - \text{SS Instar} + \text{SS Insect}$$

Two-way ANOVA

Source of variation	SS	df	MS	F	P value
Insects	426834.67	2	213417.34	67.08*	<0.05
Instars	1164912.00	3	388304.00	122.06**	<0.01
Interaction	380760.00	6	63460.00	19.95**	<0.01
Within groups	114528.00	36	3181.33		

$$\text{Total } 2087034.67 - (4 \times 3 \times 4) - 1$$

$$= 48 - 1$$

$$= 47$$

* Significant at 5% level of significance

** Significant at 1% level of significance

4. Discussion

Stav *et al.* [30] reported that the predaceous dragonfly larvae of *Anax imperator* produced 52% reduction in *Cs. longiareolata* oviposition in outdoor artificial pools. Odonata larval stages are large that requires large space and might polyphagous and might be in efficient for mosquito control besides they prefer somewhat clean non polluted aquatic habitat. Hemipteran are of sucking type, slow consumer and whole larvae are not consuming so pathogens might be present on the mosquito

larvae. This made somewhat deficient as biocontrol agent despite being able to live in both polluted and non-polluted aquatic habitats. *Lutzia* on other hand having chewing type and voracious eater. Short instar periods of 2.5 days for each moulting in *Lutzia* against 4 to 6 days of other two larval stages seems to be no effect on the efficacy on predated the mosquito larvae (Fig. 2).

Although these are successful examples of predators, there are difficulties associated with rearing; colonization and handling

which are obstacles to a more widespread utilization of predaceous aquatic insects [25]. The second difficulty is polyphagy that has advantages and disadvantages [31]. An advantage is that these predators can survive when mosquito larvae are rare or absent, while a disadvantage is that they may not reduce mosquito larvae due to availability of alternative preys. The third difficulty is the presence of other invertebrates and vertebrate predators that may reduce the abundance of the predaceous insects [32]. The fourth difficulty is predators may interfere through chemical or other cues; for instances the hydrophilid *Tropisternus lateralis* [33] and the phantom midge *Chaoborus albatrus* [34], avoid laying eggs in pools with fish. The fifth difficulty is the avoidance by mosquitoes of water containing invertebrate predators such as backswimmers and dragonflies and makes predator's impact more complicated. Lee [35] found that mosquito larvae are consumed more than pupae by predators and assumed that this was due to the inclination of pupae to exhibit rapid tumbling action when startled. Contrarily, both bugs of family belostomatidae (Order: Hemiptera) and *Toxorhynchites* mosquito larvae have an advantage over the other aquatic predaceous insects that restrict their prey selection to the larval instars only. This is worthy of note, in particular for mosquito vectors of diseases, since pupal reduction directly reduces mosquito emergence and subsequent disease transmission.

Introduction of predatory mosquito species is one of the targeted approaches for control of immature forms of mosquitoes. *Lutzia (Metalutzia) fuscana* is one of such species of mosquito whose larvae is reported to feed upon vector species larvae i.e *Anopheles*, *Aedes* and *Culex* species in several parts of India [36, 37]. Its predatory habit was found to be excellent yet non-targeted in the mesic environmental conditions due to availability of several outdoor breeding habitats. Stable breeding of *Lutzia (Metalutzia) fuscana* has been first time reported from this arid zone [38]. The genus of predatory mosquito *Lutzia fuscana* has been elevated from *Culex* to *Lutzia* (earlier *Lutzia* was subgenus) earlier *Culex (Lutzia) fuscana* (Wiedmann, 1820) is now *Lutzia (Metalutzia) fuscana* (Wiedmann) [39].

5. Conclusions

To find out the insect consuming larvae significantly more than others Turkey's HSD for 3 means and 2 means are calculated and is found to be 61.19 and 48.79. Comparing the differences between the means, all means are found to be significantly different. The insect consuming larvae significantly than others is Odonata.

In the same way, to find out the instar of the larvae at which insects consume them in significant amount Turkey's HSD is calculated for 4, 3 and 2 means and they are 79.77, 70.66 and 56.34 respectively. Comparing the differences in the means with these values, it is found that insects consume larvae at 4th instar significantly more than the larvae at other stages.

The three larvae for managing or even controlling the mosquito larvae, *Lutzia* might be the most efficient/effective candidate releasing in wild to control for it is inhabitable to the any habitats whether polluted or non-polluted but the Odonata and Hemiptera prefer somewhat non polluted aquatic habitats. The main focus of the study is to select a bio agent that has habitat boundary or constrain and nature of aquatic habitat. Hence the efficient candidate for controlling the mosquitoes of a particular habitat will be to utilize nature's

best weapon against the mosquitoes that *Lutzia tigris*.

6. Acknowledgement

We are indebted to the Principal and HOD, P. G. Department of Zoology, D M College of Science, Imphal for providing laboratory facilities. The authors are indebted to Chitralkha Devi (a student of V Sem of Zoology Department) for collecting and guiding the species of *Lutzia tigris* from Urembam. The authors are also thankful to the Ministry of Science and Technology, Department of Biotechnology, GOI for the financial assistance under No. BT/IN/Indo-US/Foldscope/39/2015 dated 20/03/2018.

7. References

1. Norbert Becker, Dušan Petric, Marija Zgomba Clive Boase, Christine Dahl, Minoo Madon, Achim Kaiser. Mosquitoes and Their Control. Springer, 2010.
2. Shaalan Essam Abdel-Salam, Deon Canyon V. Aquatic insect predators and mosquito control. Tropical Biomedicine. 2009; 26(3):223-261.
3. Chandra G, Bhattacharjee I, Chatterjee SN, Ghosh A. Mosquito control by larvivorous fish. Indian Journal of Medical Research. 2008; 127:13-27.
4. Dua VK, Pandey AC, Rai S, Dash AP. Larvivorous activity of *Poecilia reticulata* against *Culex quinquefasciatus* larvae in a polluted water drain in Hardwar, India. Journal of American Mosquito Control Association. 2007; 23:481-483.
5. Biswas D, Dutta RN, Ghosh SK, Chatterjee KK, Hati AK. Breeding habits of *Anopheles stephensi* Liston in an area of Calcutta. Indian Journal Malariology. 1992; 29(3):195-198.
6. Chandras RK, Rajagopalan PK. Observations on mosquito breeding and the natural parasitism of larvae by a fungus *Coelomomyces* and a mermithid nematode *Romanomermis* in paddy fields in Pondicherry. Indian Journal of Medical Research. 1979; 69:63-67.
7. Poopathi S, Tyagi BK. The Challenge of Mosquito Control Strategies: from Primordial to Molecular Approaches. Biotechnology and Molecular Biology Review. 2006; 1(2):51-65.
8. Mattingly PF. The culicine mosquitoes of the Indomalayan Area. Part V: genus *Aedes* Meigen, subgenera *Mucidus* Theobald, *Ochlerotatus* Lynch Arribalzaga and *Neome laniconion* Newstead. Brit Mus Nat Hist London, 1961, 62.
9. Ikeshoji T, Mulla MS. Oviposition attractants for four species of mosquitoes in natural breeding waters. Ann Ent Soc Am. 1970; 63(5):1322-1327.
10. Panicker KN, Bai MG, Sabesan S. A note on laboratory colonization of *Culex (Lutzia) fuscana* Wiedemann, 1820 (Diptera: Culicidae). Indian Journal of Medical Research. 1982; 75:45-46.
11. Ramalingam S, Ramakrishnan K. Re-description of *Aedes (Alanstonea) Brevitibia* (Edwards) from Brunei, Borneo. Proceedings of Entomological Society Washington. 1971; 73:231-238.
12. Kumar R, Hwang JS. Larvicidal efficacy of aquatic predators: A perspective for mosquito control. Zoological Studies. 2006; 45(4):447-466.
13. Bay EC. Predatory-prey relationships among aquatic insects. Annual Review of Entomology. 1974; 19:441-453.

14. Shaalan EA. Integrated control of two mosquito species *Aedes aegypti* and *Culex annulirostris*. Ph. D. Thesis. South Valley University, Egypt, 2005.
15. Shaalan EA, Canyon DV, Reinhold M, Yones WFM, Abdel-Wahab H, Mansour A. A mosquito predator survey in Townsville, Australia and an assessment of *Diplonychus* sp. and *Anisops* sp. predatorial capacity against *Culex annulirostris* mosquito immatures. *Journal of Vector Ecology*. 2007; 32(1):16-21.
16. Legner EF. Biological control of Diptera of medical and veterinary importance. *Journal of Vector Ecology*. 1994; 20(1):59-120.
17. Rodriguez-Castro VA, Quiroz-Martinez H, Solis-Rojas H, Tejada LO. Mass rearing and egg release of *Buenoa scimitra* Bare as biocontrol of larval *Culex quinquefasciatus*. *Journal of the American Mosquito Control Association*. 2006; 22(1):123-125.
18. El-Rayah E. Dragonfly nymphs as active predators of mosquito larvae. *Mosquito News*. 1975; 35:229-230.
19. Sebastian A, Thu MM, Kyaw M, Sein MM. The use of dragonfly nymphs in the control of *Aedes aegypti*. *Southeast Asian Journal of Tropical Medicine and Public Health*. 1980; 11(1):104-107.
20. Sebastian A, Sein MM, Thu MM, Corbet PS. Suppression of *Aedes aegypti* (Diptera: Culicidae) using augmentative release of dragonfly larvae (Odonata: Libellulidae) with community participation in Yangon, Myanmar. *Bulletin of Entomological Research*. 1990; 80:223-232.
21. Chatterjee SN, Ghosh A, Chandra G. Eco-friendly control of mosquito larvae by *Brachytron pratense* nymph. *Journal of Environmental Health*. 2007; 69(8):44-49.
22. Mandal SK, Ghosh A, Bhattacharjee I, Chandra G. Biocontrol efficiency of odonate nymphs against larvae of the mosquito, *Culex quinquefasciatus* Say, 1823. *Acta Tropica*. 2008; 106:109-114.
23. Breene RG, Sweet MH, Olson JK. Analysis of the gut contents of naiads of *Enallagma civile* (Odonata: Coenagrionidae) from a Texas pond. *Journal of the American Mosquito Control Association*. 1990; 6(3):547-548.
24. Collins LE, Blackwell A. The biology of *Toxorhynchites* mosquitoes and their potential as biocontrol agents. *Biocontrol News and Information*. 2000; 21:105-116.
25. Garcia R. Arthropod predators of mosquitoes. *Bulletin of the Society of Vector Ecology*. 1982; 7:45-47.
26. Pal R, Ramalingam S. Invertebrate predators of mosquitoes. *World Health Organization WHO/VBC/81*, 1981, 799.
27. Lacey LA, Orr BK. The role of biological control of mosquitoes in integrated vector control. *American Journal of Tropical Medicine and Hygiene*. 1994; 50(6):97-115.
28. Mogi M. Insects and other invertebrate predators. *Journal of the American Mosquito Control Association*. 2007; 23(3):93-109.
29. Quiroz-Martine H, Rodriguez-Castro A. Aquatic insects as predators of mosquito larvae. *Journal of the American Mosquito Control Association*. 2007; 23(2):110-117.
30. Stav G, Blaustein L, Margalith J. Experimental evidence for predation risk sensitive oviposition by a mosquito, *Culiseta longiareolata*. *Ecological Entomology*. 1999; 24:202-207.
31. Murdoch WW, Scott MA, Ebsworth P. Effects of the general predator, *Notonecta* (Hemiptera) upon a fresh water community. *Journal of Animal Ecology*. 1984; 53:791-808.
32. Larson DJ. Odonate predation as a factor influencing dytiscid beetle distribution and community structure. *Questiones Entomology*. 1990; 26:151-162.
33. Resetarits WJ. Colonization under threat of predation: avoidance of fish by an aquatic beetle, *Tropisternis lateralis* (Coleoptera: Hydrophilidae). *Oecologia*. 2001; 129:155-160.
34. Petranka JW, Fakhoury K. Evidence of a chemically-mediated avoidance response of ovipositing insects to bluegills and green frog tadpoles. *Copeia*. 1991; 1:234-239.
35. Lee FC. Laboratory observations on certain mosquito larval predators. *Mosquito News*. 1967; 27(3):332-338.
36. Geetha Bai, Viswam MK, Panick KN. *Culex (Lutzia) fuscus* (Diptera: Culicidae) - a predator of mosquito. *Indian Journal of Medical Research*. 1982; 76:837-839.
37. Panicker KN, Bai MG, Sabesan S. A note on laboratory colonization of *Culex (Lutzia) fuscus* Wiedemann, 1820 (Diptera: Culicidae). *Indian Journal of Medical Research*. 1982; 75:45-46.
38. Singh H, Marwal R, Mishra A, Singh KV. Invasion of the predatory mosquito *Culex (Lutzia) fuscus* in the western desert parts of Rajasthan, India. *Poland Journal of Entomology*. 2013; 82:49-58.
39. Tanaka K. Studies on the pupal mosquitoes of Japan (9). Genus *Lutzia*, with establishment of two new subgenera, *Metalutzia* and *Insulalutzia* (Diptera, Culicidae). *Japanese Journal of Systematic Entomology*. 2003, 159-169.