

## International Journal of Mosquito Research

ISSN: 2348-5906  
CODEN: IJMRK2  
IJMR 2018; 5(6): 39-42  
© 2018 IJMR  
Received: 10-09-2018  
Accepted: 15-10-2018

**Sundara Mahalingam Balaji**  
Post-Graduate Department of  
Biotechnology Ayya Nadar  
Janaki Ammal College  
(Autonomous), Sivakasi,  
Tamil Nadu, India

**Kani Manjula**  
Post-graduate Department of  
Biotechnology Ayya Nadar  
Janaki Ammal College  
(Autonomous), Sivakasi,  
Tamil Nadu, India

# Biosynthesis of silver nano composite using agro waste and its larvicidal properties

**Sundara Mahalingam Balaji and Kani Manjula**

### Abstract

Mosquito has attracted considerable attention in the medical and social regions of the world. They are the vectors of many diseases accounting for huge mortality and morbidity worldwide. In the present experiment we prepared a silver nano composite using paddy straw and tested its potential larvicidal activities against *Aedes aegypti* and *Anopheles gambiae* larvae at 25, 50, 100 ppm concentration. The synthesised nanoparticles using paddy straws were characterised by Ultra Violent resonance, Fourier transform infrared spectroscopy, Scanning Electron Microscopy and Energy-dispersive X-ray spectroscopy analysis. In larvicidal assay, the fourth instar larvae are exposed to different concentrations of silver nano composite aqueous extract for 24 hours. From the experiment it is revealed that the mean concentration to kill 50% of larval population is found to be 13.625 ppm for *Aedes aegypti* and 12.270 ppm for *Anopheles gambiae*.

**Keywords:** Silver nano composite, larvicidal, *Aedes aegypti* and *Anopheles gambiae*

### 1. Introduction

Mosquitoes widely transmit pathogens that spread endemic diseases among human beings and animals. It is reported that approximately 1.5 million new cases of malaria are recorded every year in India and it also ends up in nearly 1000 deaths<sup>[1]</sup>. According to 2006–2007 survey in India regarding the epidemics of mosquito-borne diseases concluded 1.4 million suspected and 2000 confirmed cases of Chikungunya virus and 1000 deaths from Japanese encephalitis<sup>[2]</sup>. So it is high time for Vector control by developing new and improved mosquito control methods that are economical and effective. The vector control methods should also be safe for non-target organisms and the environment. Nanoparticles are recognised for their unique optical properties. The metals like gold and silver have a broad absorption band in the visible region of the electromagnetic spectrum<sup>[3, 4]</sup>. The properties of these metals are used in advanced medicine, opto-electronics, chemical catalysis, sensors, drug delivery, and for etching and cutting as they are able to change their shape, size, and the surrounding medium<sup>[5,6,7,8,9,10]</sup>.

To escape from the biting of mosquitoes people must use repellents and target larval stage of mosquito as it would be the better strategy to lower the incidence of mosquito-transmitted diseases. In larval stage have less mobility in breeding habitat, so planning control measures at this stage is comparatively easy<sup>[11]</sup>. The present practice to control mosquito larvae is the use of insecticides like carbamate, organophosphate and pyrethroids. Mosquitoes died for insecticides but the prolonged usage of it made mosquitoes get resistance towards insecticides<sup>[12]</sup>. Mosquitoes got varying amount of resistance to commonly used insecticides like temephos, fenthion, malathion and Dichloro Diphenyl Trichloroethane (DDT)<sup>[13]</sup>. Moreover, chemical insecticides remain a threat to non-target species, their environment as well as their food chain<sup>[14]</sup>. Considering this, insect control agents from biological sources would always be considered as a safe and effective way.

Several plants like *Azadirachta indica*<sup>[15]</sup>, *Aloe vera*<sup>[16]</sup>, *Plumeria rubra*<sup>[17]</sup>, *Nelumbo nucifera*<sup>[18]</sup> and *Emblica officinalis*<sup>[19]</sup>, *Medicago sativa* sprouts<sup>[20]</sup> are screened successfully for silver nanoparticles synthesis by various researchers. Nowadays, phytosynthesised silver nanoparticles as a mosquito larvicidal agent are gaining importance instead of chemical insecticide. This is because of their less harmful effect to non-targeted species and novelty in mechanism of action<sup>[21]</sup>. It is our dire need to control vector borne diseases as they are the major public health concern at global level.

### Correspondence

**Sundara Mahalingam Balaji**  
Post-Graduate Department of  
Biotechnology Ayya Nadar  
Janaki Ammal College  
(Autonomous), Sivakasi,  
Sivakasi, Tamil Nadu, India

Once a person is infected with malaria, it involves typical medical treatment and there is report of resurgence of malaria after eradication in many countries [10]. Hence the present study is undertaken to synthesize a nanocomposite using agricultural paddy straw and screen its larvicidal properties against *Aedes aegypti* and *Anopheles gambiae*.

## 2. Materials and Methods

### 2.1 Collection of plant materials and preparation of paddy straw extract

Fresh and healthy straw are collected locally and rinsed thoroughly with tap water followed by distilled water to remove all the dust and unwanted visible particles and are placed in a shadow and dried well. Then they were cut into small pieces and ground well to get a powder form.

### 2.2 Synthesis, characterization, and formulation of silver nanoparticles

10ml of paddy straw extract was added into 90 ml of aqueous solution of 1mM silver nitrate ( $AgNO_3$ ) for the reduction of silver nitrate and kept in a dark room at 37 °C for 24 hours. After 24 hours, colour of the solution changed from green to dark brown indicating the formation of silver nanoparticles. The bioreduced silver nanoparticles solution was measured using UV- Visible absorbance spectrophotometer modified method of Joyti and Singh [22]. The surface group of the nanoparticles is qualitatively confirmed by using Fourier transform infrared (FTIR) spectroscopy, X-ray diffraction (XRD) to determine the crystalline structure and visible observation by Scanning Electron Microscopy.

### 2.3 Collection and rearing of Mosquito larvae

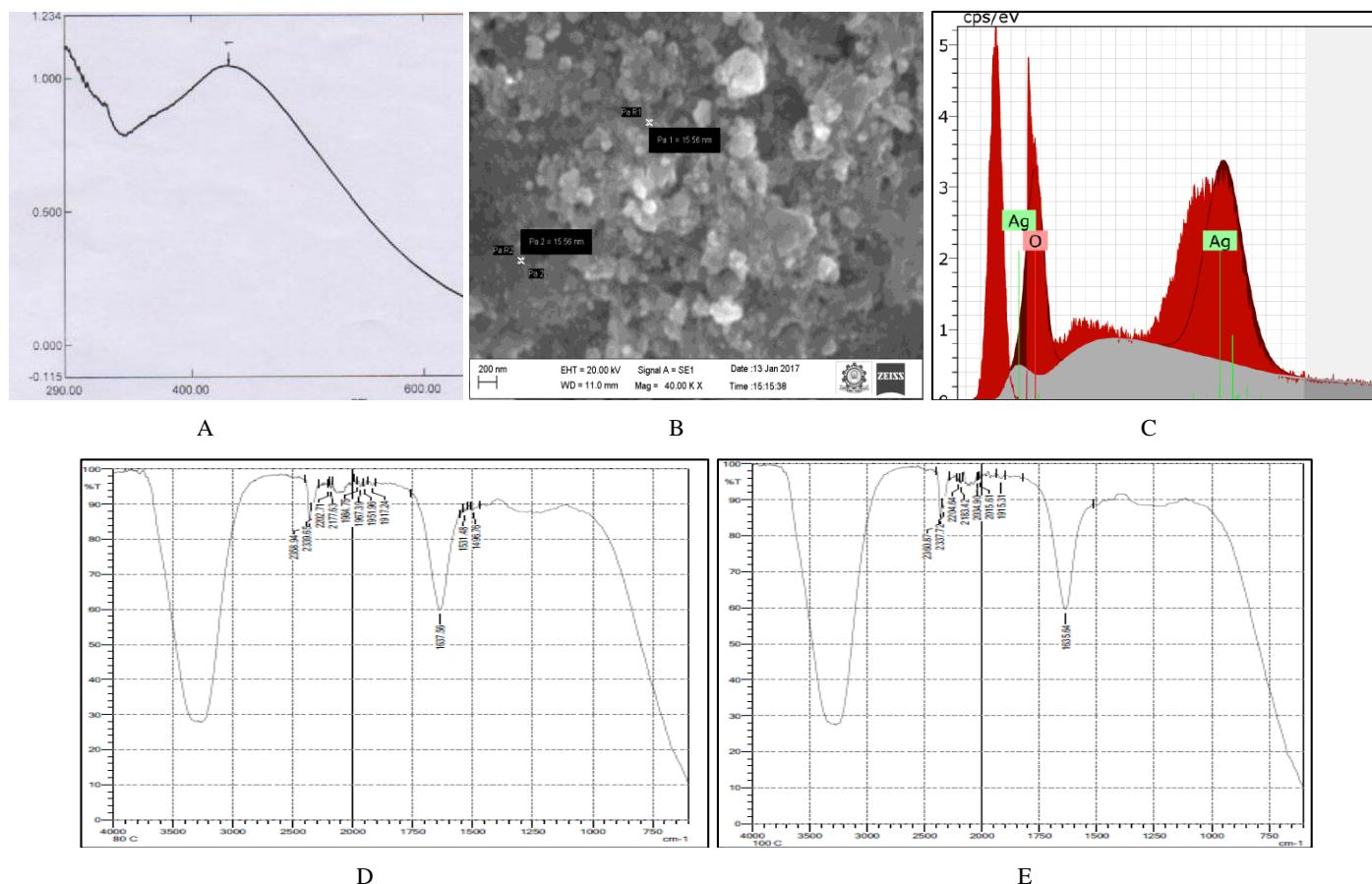
Larval strains of *Aedes aegypti* and *Anopheles gambiae* are collected from the Government research Institute of Indian Council for Medical Research, Madurai. Larvae are maintained in enamel tray containing dechlorinated tap water mixed with preparation of dog biscuits and yeast extracts under suitable conditions.

### 2.4 Mosquito larvicidal bioassay

For bioassay test, IV<sup>th</sup> instar larvae of *Aedes aegypti* and *Anopheles gambiae* are taken. The larvicidal activity is studied by the standard procedure recommended by WHO [23]. Green synthesized AgNPs are prepared into different concentrations of 25, 50 and 100 ppm with double distilled water. Twenty-five reared early fourth instar stage larvae are transferred by means of the dropper to (100 mL beakers) each concentration and the experiment is performed in three replicates under the laboratory conditions. Mortality of the larvae is calculated at 12 and 24 hr of exposure periods. During the both exposure periods, no food is supplied to the larvae and the percentage of mortality was calculated. The lethal concentrations in parts per million  $LC_{50}$  are also calculated using Probit analysis [24].

## 3. Results

Silver particles which are the primary result of this experiment changed its colour in various concentrations and the aqueous solution of silver ion complex started to change the colour from yellowish brown to dark brown. The synthesis of silver nanoparticles in the solution is further analysed and confirmed by UV-visible spectroscopy. The resonance is observed after 24 hrs at 431nm for the green synthesis.



**Plate 1:** Observations of synthesised nanoparticle (A – UV light absorbance; B – Scanning electron micrograph; C – XRD analysis; D&E – FTIR peaks)

The silver composite prepared in this investigation is found to exhibit uniform sized cubic grains with a range of 15.56 nm particles. The presence of silver particles was also quantified using XRD pattern and obtained a peak at 3 keV. The bioreduction of silver nanoparticles is further confirmed by FTIR spectra of aqueous silver nanocomposite prepared from the paddy straw extract show transmittance peaks at 1496.76, 1637.56, 1917.24, 2202.71, 2339.65 /cm. These peaks indicate that the carbonyl group formed amino acid residues and that these residues capped the silver nanoparticles to prevent agglomeration, thereby stabilizing the medium. During the bioreduction there is a disappearance of many peaks due to the structural change in paddy straw. Different concentrations of 25, 50 and 100 ppm silver nano composite using paddy straw are prepared and observed for mortality of mosquito larvae of *Aedes aegypti* and *Anopheles gambiae*. From the experiment it is found that maximum number of mortality is found to be present in the concentration of 30 ppm in both the experimental larvae when compared to other concentrations (Table 1). No mortality is recorded in control sample. Synthesized nano particles from paddy extract are more active against the *Aedes aegypti* species presenting LC<sub>50</sub> value of 13.625 when compared to LC<sub>50</sub> value of 12.270 *Anopheles gambiae*.

#### 4. Discussion

The typical Silver nanoparticles spectrum has a maximum absorption in the range of 420-450 nm. This absorption is a unique property of metal nanoparticles called SPR (Surface Plasmon Resonance) that arise due to conduction of electrons on the surface of silver nanoparticles. SPR for different metal nanoparticles synthesised using different substances are reported in previous studies such as, for gold nanoparticles it is around 540 nm Das *et al.* [25] and 315 nm for Zinc sulphide (ZnS) nanoparticles Hudlikar *et al.* [26]. In the present study the absorption spectrum is found to be 431 nm. In the present study, the larvicidal activity of silver nitrate nanoparticle may be due to the direct attack mechanism on the larva and is supported by Naresh *et al.* [27] who used silver nanoparticles for the control of mosquito causing diseases like Malaria, dengue and filariasis. While comparing the prokaryotic systems, the silver nano particles have multiple targets for biocidal activity by causing structural damages to the organism Kim *et al.* [28], generation of reactive oxygen species and interfering with DNA replication (Liau *et al.* [29] and Feng *et al.* [30]. Administration of silver nanoparticles *in vivo* to Zebra fish embryos by Lee *et al.* [31] increased deformation rates and ultimately led to death. The mechanism which causes the death of the larvae could be the ability of the nanoparticles to penetrate through the larval membrane. The silver nanoparticles in the intracellular space can bind to sulphur containing proteins or to phosphorus containing compounds like DNA, leading to the denaturation of some organelles and enzymes according to Rai *et al.* [32]. The mortality effect of silver nanoparticles on mosquito larvae may be enabled by the small size of the particles, which allows passage through the insect cuticle and into individual cells where they interfere with molting and other physiological processes.

#### 5. Conclusion

From the experiment it is revealed that low concentration of the prepared nano composite can effectively kill the larvae of

the mosquito but still field study is essential to confirm the activity. In the future, the mode of action of nanoparticles on the larvae can also be studied using latest methodologies.

#### 6. Acknowledgement

The authors are grateful to The Management of Ayya Nadar Janaki Ammal College, Sivakasi, having provided us the necessary lab facilities to do the work in the Department of Biotechnology.

**Table 1:** Larvicidal activity of synthesized silver nanoparticles using paddy straw leaf extract against *Aedes aegypti* and *Aedes gambiae* larvae.

Mosquito species	Concentration (ppm)	Mortality	LC <sub>50</sub>
<i>Aedes aegypti</i>	Control	0	13.625
	25	3±1.52	
	50	13±2.00	
	100	22.67±1.52	
<i>Anopheles gambiae</i>	Control	0	12.270
	25	3.67±1.52	
	50	17.33±2.51	
	100	23.66±1.52	

#### 7. References

- Kumar A, Valecha N, Jain T, Dash AP. Burden of malaria in India: retrospective and prospective view. *American J Trop. Med. Hyg.* 2007; 77:69-78.
- Laneri K, Bhadra A, Ionides EL, Bouma M, *et al.* Forcing versus feedback: Epidemic malaria and monsoon rains in Northwest India. *PLoS Comput. Biol.* 2010; 9:1-13.
- Kreibig C, Vollmer HF. *Optical Properties of Metal Clusters*. Springer Series in Inorganic Chemistry 15. Berlin: Springer, 1995.
- Mulvaney P. Surface plasmon spectroscopy of nanosized metal particles. *Langmuir.* 1996; 12:788-800.
- Che M, Bennett CO. The influence of particle size on the catalytic properties of supported metals. *Adv Synthesis Catalysis.* 1989; 36:55-172.
- Elghanian R, Storhoff JJ, Mucic RC, Letsinger RL, Mirkin CA. Selective colorimetric detection of polynucleotides based on the distance dependent optical properties of gold nanoparticles. *Science.* 1997; 277:1078-1081.
- El-Sayed IH, Huang X, El-Sayed MA. Selective laser photothermal therapy of epithelial carcinoma using anti-EGFR antibody conjugated gold nanoparticles. *Cancer Lett.* 2006; 239:129-135.
- Aurel Y, Jan G, Paul VL, Thijs W, *et al.* Fast, ultrasensitive virus detection using a young interferometer sensor. *Nano Lett.* 2007; 7:394-397.
- Jain PK, El-Sayed IH, El-Sayed MA. Au nanoparticles target cancer. *Nano Today.* 2007; 2:18-29.
- Cohen JM, Smith DL, Cotter C, Ward A, Yamey G. Malaria resurgence: A systematic review and assessment of its causes. *Malar. J.* 2012; 11:122.
- Howard AF, Zhou G, Omlin FX. Malaria mosquito control using edible fish in western Kenya: preliminary findings of a controlled study. *BMC Public Health.* 2007; 7:199.
- Raghvendra K. Chemical insecticides in malaria vector

- control in India. ICMR Bulletin. 2012, 32.
13. Tikar SN, Mendki MJ, Chandel K, Parashar BD, Prakash S. Susceptibility of immature stages of *Aedes* (*Stegomyia*) *aegypti*; vector of dengue and chikungunya to insecticides from India. Parasitology Res. 2008; 102:907-913.
  14. Sarwar M, Ahmad N, Toufiq M. Host plant resistance relationships in chickpea (*Cicer arietinum* L.) against gram pod borer (*Helicoverpa armigera* Hubner). Pak. J Bot. 2009; 41:3047-3052.
  15. Shankar SS, Rai A, Ahmad A, Sastry M. Rapid synthesis of Au, Ag, and bimetallic Au core-Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. J Colloid Interface Sci. 2004; 275:496-502.
  16. Chandran SP, Chaudhary M, Pasricha R, Ahmad A, Sastry M. Synthesis of gold nanotriangles and silver nanoparticles using *Aloe vera* plant extract. Biotechnol Prog. 2006; 22:577-583.
  17. Patil CD, Patil SV, Borase HP, Salunke BK, Salunkhe RB. Larvicidal activity of silver nanoparticles synthesized using *Plumeria rubra* plant latex against *Aedes aegypti* and *Anopheles stephensi*. Parasitology Res. 2012; 110:1815-1822.
  18. Santhoshkumar T, Rahuman AA, Rajakumar G, Marimuthu S, Bagavan A, et al. Synthesis of silver nanoparticles using *Nelumbo nucifera* leaf extract and its larvicidal activity against malaria and filariasis vectors. Parasitology Res. 2011; 108:693-702
  19. Ankanwar B, Damle C, Ahmad A, Sastry M. Biosynthesis of gold and silver nanoparticles using *Embllica officinalis* fruit extract, their phase transfer and transmetallation in an organic solution. J Nanosci Nanotechnol. 2005; 5:1665-1671.
  20. Gardea-Torresdey JL, Gomez E, Peralta-Videa JR, Parsons JG, Troiani H, et al. *Alfalfa* sprouts: A natural source for synthesis of silver nanoparticles. Langmuir. 2003; 19:1357-1361.
  21. Marimuthu S, Rahuman AA, Rajakumar G, Santhoshkumar T, Kirthi AV, et al. Evaluation of green synthesized silver nanoparticles against parasites. Parasitology Res. 2011; 108:1541-1549.
  22. Jyoti K, Singh A. Green synthesis of nanostructured silver particles and their catalytic application in dye degradation. Journal of Genetic Engineering and Biotechnology. 2016; 14:311-317.
  23. WHO Guidelines for Laboratory and Field Testing of Mosquito Larvicides, World Health Organization document WHO, 2005.
  24. Bhuvaneshwari R, Xavier RJ, Arumugam M. Larvicidal property of green synthesized silver nanoparticles against vector mosquitoes (*Anopheles stephensi* and *Aedes aegypti*). Journal of King Saud University – Science. 2016; 28:318-323
  25. Das SK, Das AR, Guha AK. Gold nanoparticles: microbial synthesis and application in water hygiene management. Langmuir. 2009; 25:8192-8199.
  26. Hudlikar M, Joglekar S, Dhaygude M, Kodam K. Latex-mediated synthesis of ZnS nanoparticles: green synthesis approach. J Nanopart Res. 2012; 14:865.
  27. Naresh KA, Kadarkarai M, Chandrababu R, Pari M, Donald RB. Green Synthesis of Silver Nanoparticles for the Control of Mosquito Vectors of Malaria, Filariasis, and Dengue. Vector-Borne and Zoonotic Diseases. 2012; 12(3):262-268.
  28. Kim JS, Kuk E, Yu KN, Kim JS, et al. Antimicrobial effects of silver nanoparticles. Nanomed Nanotechnol. Biol. Med. 2007; 3:95-101.
  29. Liao SY, Read DC, Pugh WJ, Furr JR, Russell AD. Interaction of silver nitrate with readily identifiable groups: relationship to the antibacterial action of silver ions. Lett. Appl. Microbiol. 1997; 25:279-283.
  30. Feng QL, Wu J, Chen GQ, Cui FZ, Kim TN, Kim JO. A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. J Biomed. Mater. Res. 2000; 52:662-668.
  31. Lee KJ, Nallathamby PD, Browning LM, Osgood CJ, Nancy XH. *In vivo* imaging of transport and biocompatibility of silver nanoparticles in early development of zebrafish embryos. ACS Nano. 2007; 1:133-143.
  32. Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. Biotechnol. Adv. 2009; 27:76-83.